

PEST CONTROL UNDER DIFFERENT LIGHT CONDITIONS

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Hochschule Geisenheim University
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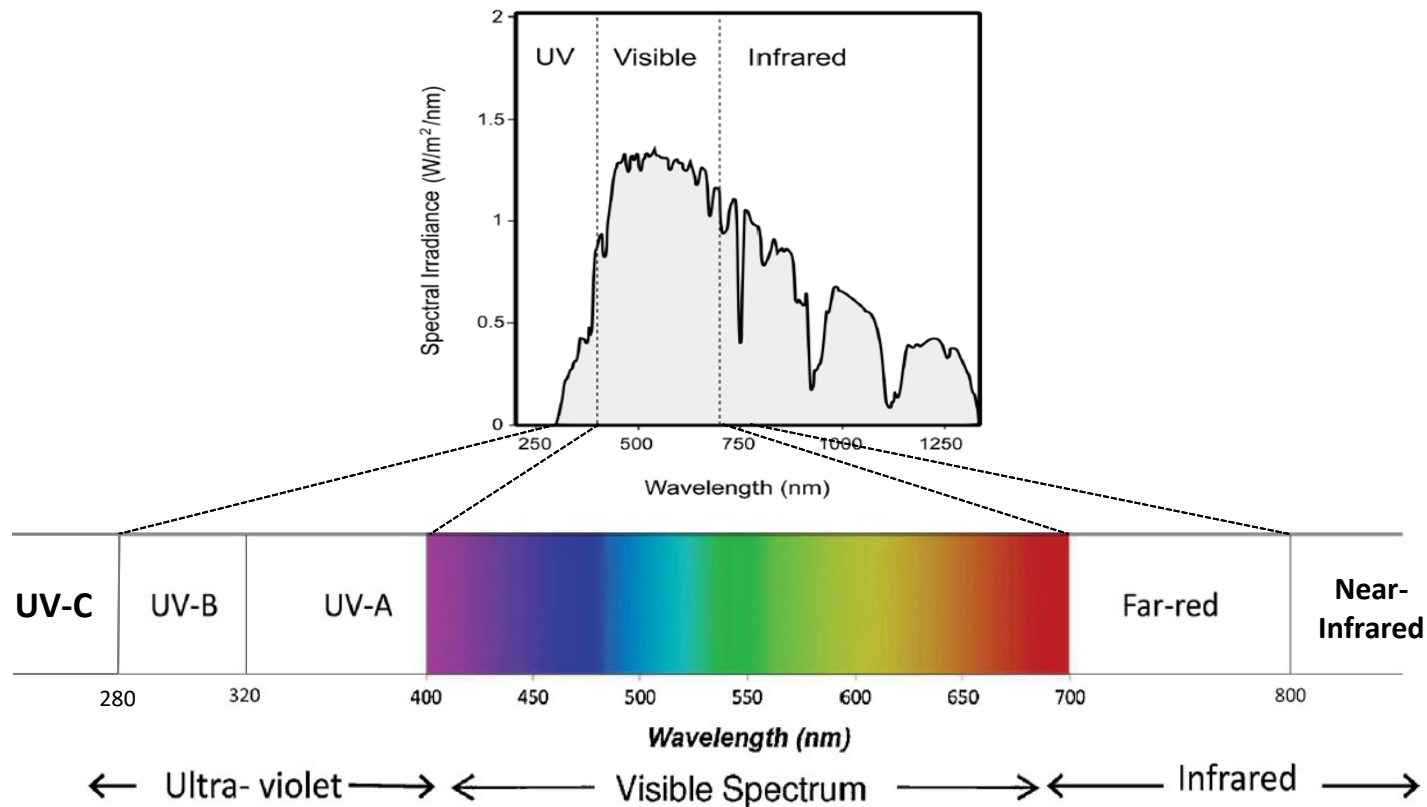
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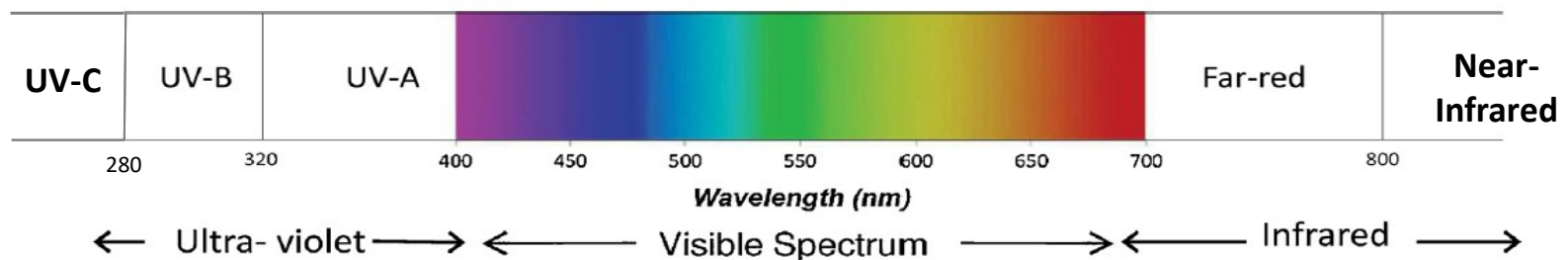
- **What is light?**

electromagnetic radiation within a certain portion of the electromagnetic spectrum



Greenhouse cladding changes the quality and quantity of the transmitted radiation, especially regarding:

- intensity (quantity)
- UV
- NIR
- blue wavelengths



PEST CONTROL UNDER DIFFERENT LIGHT CONDITIONS

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- **What is a pest?**

Merriam-Webster dictionary: “a plant or animal detrimental to humans or human concerns (such as agriculture or livestock)”

→ *here*: focus on insect pests in agriculture; **food competitors to humans**





- Feeding on plant sap: aphids

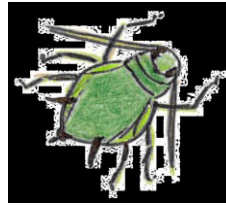




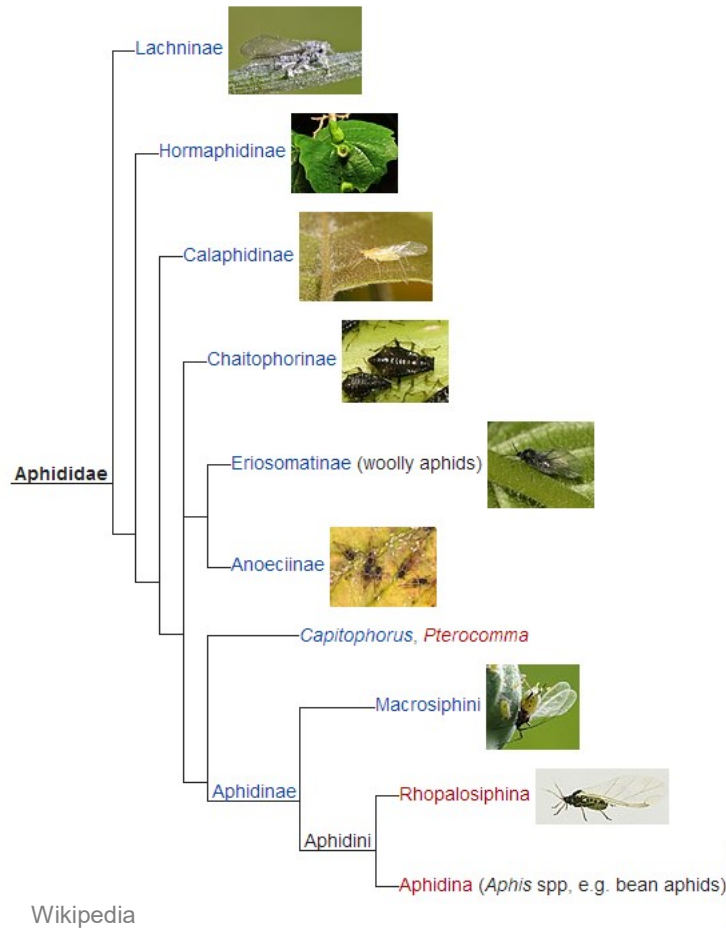
- Feeding on plant sap: aphids

Wikipedia, Jacopo Werther





- aphids: high taxonomic diversity



Many species with diverse shapes, colors and sizes!



Andreas Eichler



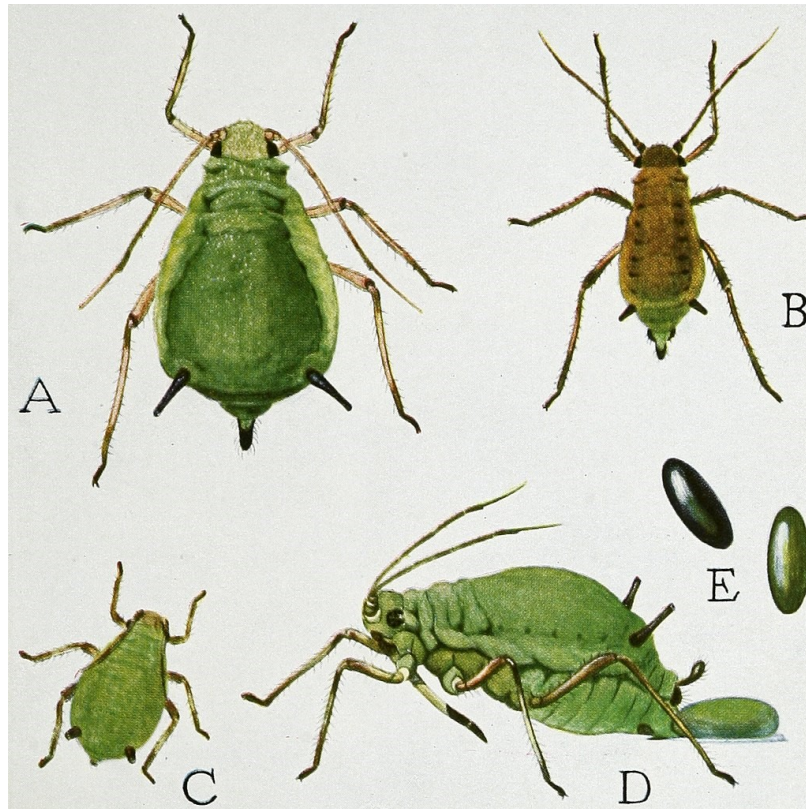
Sanjay Arachya



Alvesgaspar



- aphid life cycle, reproduction



Green apple aphid (*Aphis pomi*)
(Robert Evans Snodgrass, 1930)

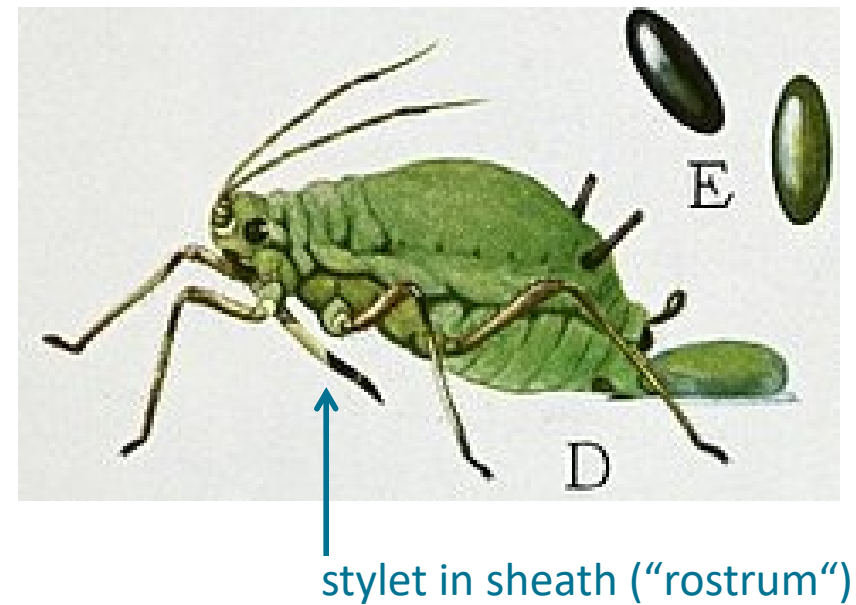
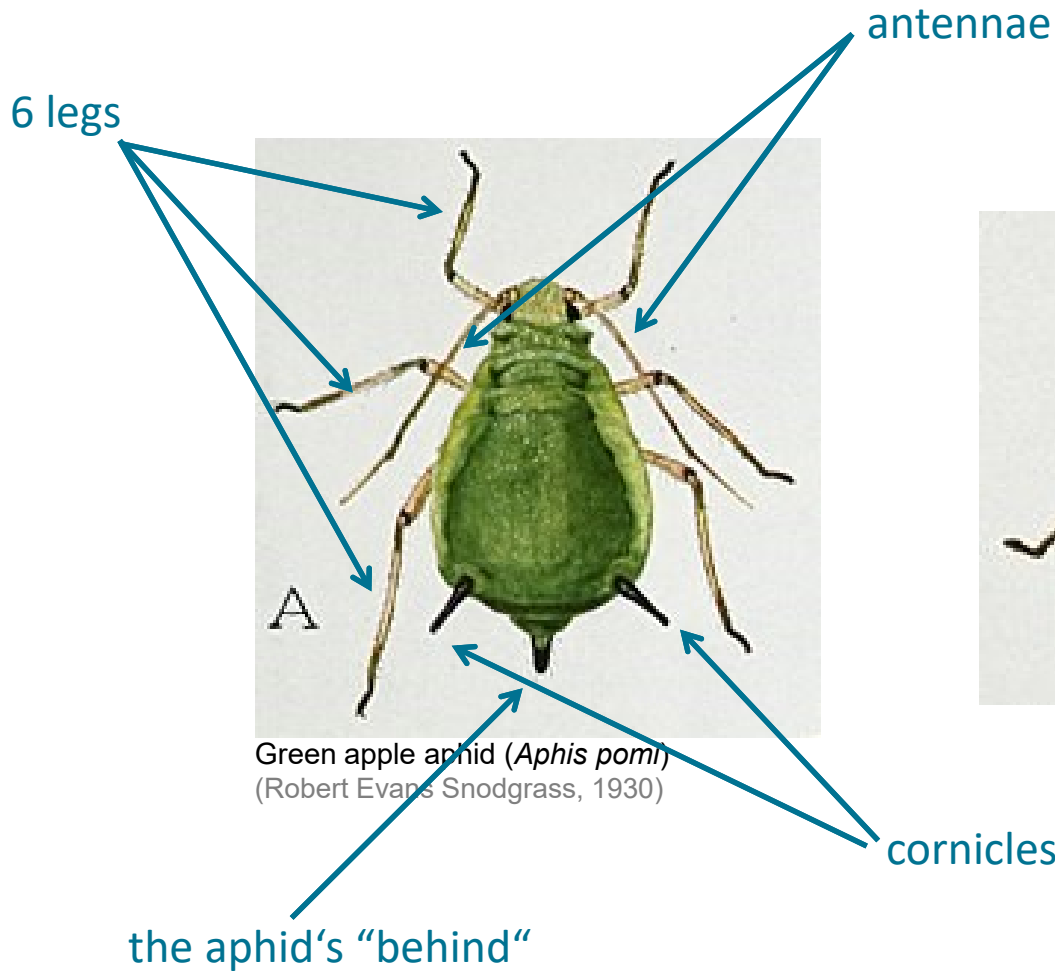
- Very diverse reproduction cycles!
- 1 common model:
 - beginning of the season: only females, asexual reproduction
 - declining food quality: winged females produced, to change host plant
 - end of the season: production of males, sexual reproduction

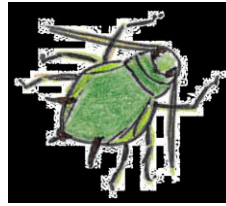


Yerpo



- aphid morphology

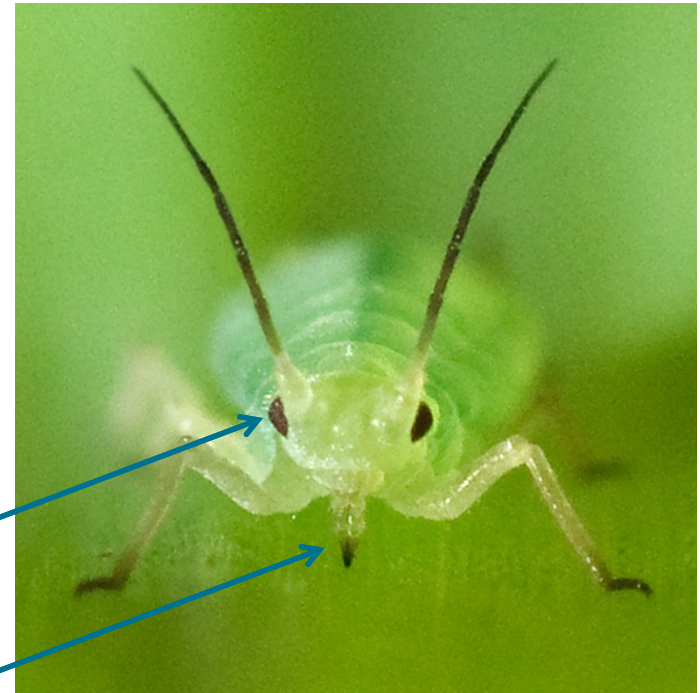




- aphid feeding



Green apple aphid (*Aphis pomi*)
(Robert Evans Snodgrass, 1930)



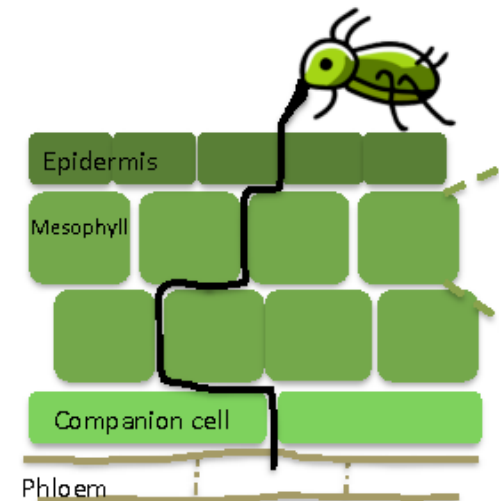
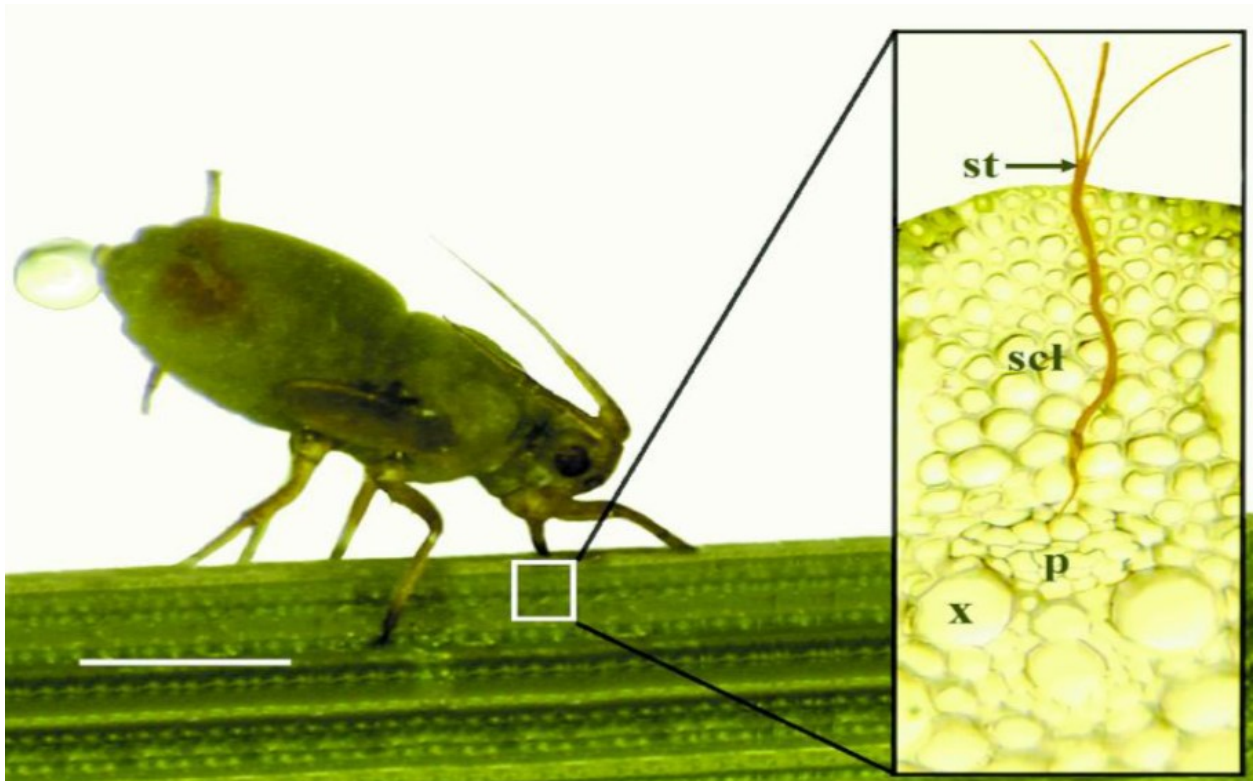
compound eye

stylet in sheath ("rostrum")

Front view of wheat aphid, *Schizaphis graminum*, showing the piercing-sucking mouthparts



- Feeding on plant sap: aphids



(Mehrabi 2016, modified)

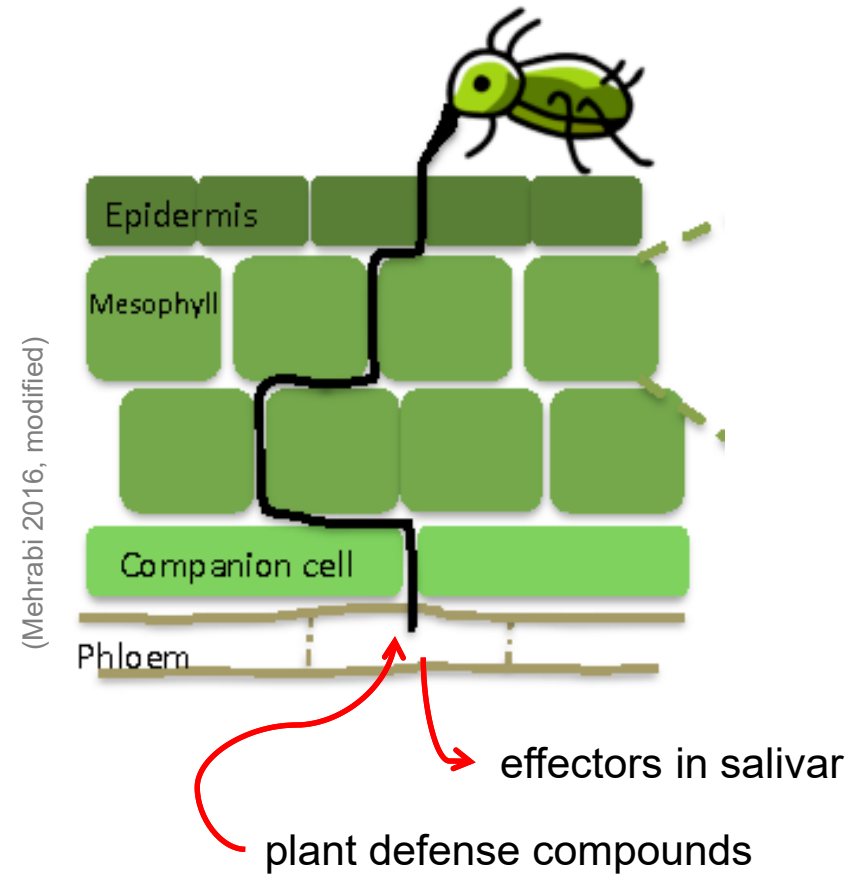
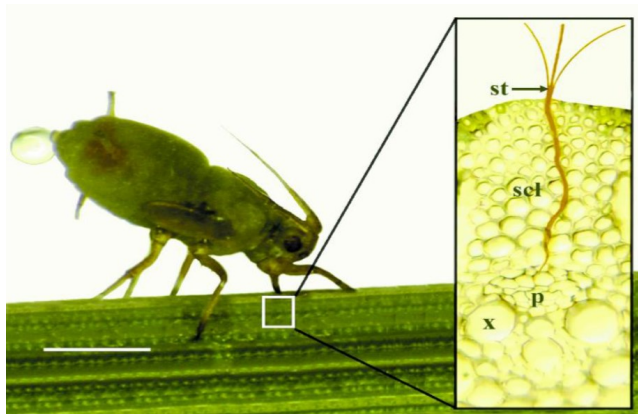
Aphid feeding on phloem sap.

st: stylet, scl: sclerenchyma, p: phloem, x: xylem. Scale bar = 1 mm.

(“Plants in Action”, Australian Society of Plant Scientists)



- Feeding on plant sap: aphids





- aphids: feeding on plant sap, excreting fluids



Sanjay Arachya

Cornicles: excretion of **defense fluid**



Amada44

Anus: excretion of **honeydew**



- also feeding on plant sap: whitefly



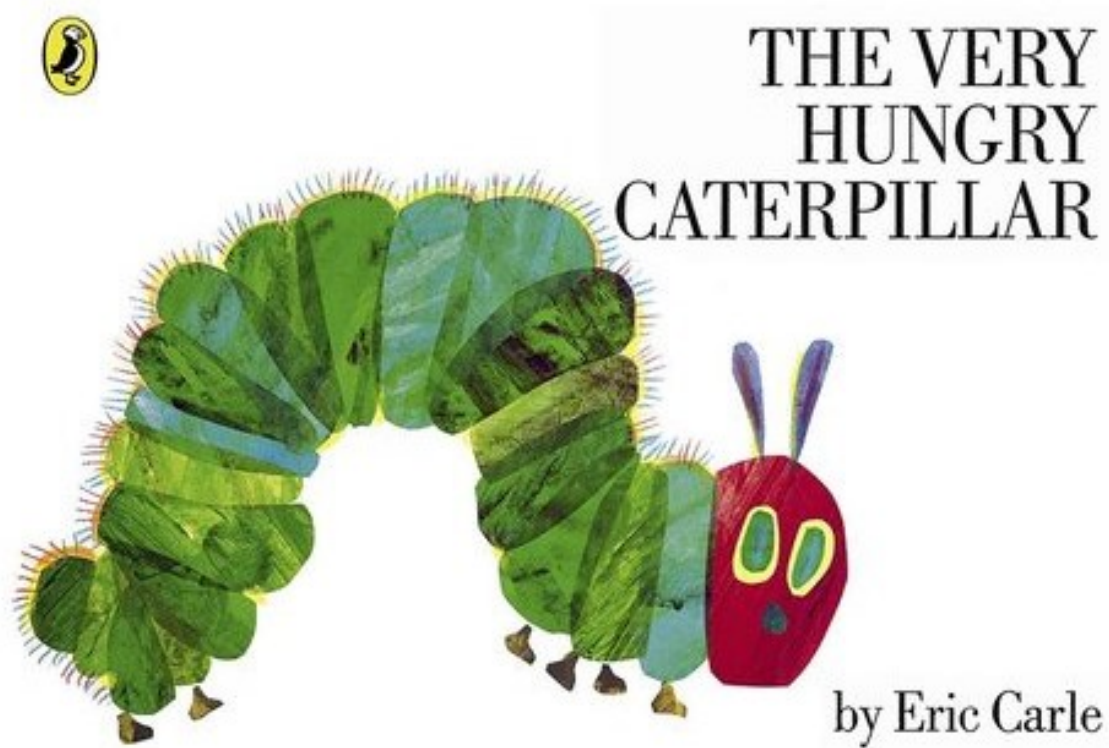
gaucho

Trialeurodes vaporariorum, Greenhouse Whitefly

- weaken plants through salivary toxins
- often spread viruses while feeding (like aphids)
- excrete honeydew that fosters mold (like aphids)
- Prominent pests:
 - *Bemisia tabaci*, Silverleaf whitefly
 - *Aleyrodes proletella*, Cabbage Whitefly
 - *Trialeurodes vaporariorum*, Greenhouse Whitefly



- Eating whole leaves: caterpillars, larvae of Lepidoptera (butterflies and moths)





- Eating whole leaves: caterpillars, larvae of Lepidoptera (butterflies and moths)



© Christoph Hoyer

Bright-line brown-eye (*Lacanobia oleraceae*)



- Eating whole leaves: caterpillars, larvae of Lepidoptera (butterflies and moths)



<http://www.kreisblatt.de/lokales/main-taunus-kreis/Gespinstmotte-spinnt-eine-Fressfeinde;art676,2990551>

Ermine moths, Yponomeutidae family

Blend5,6





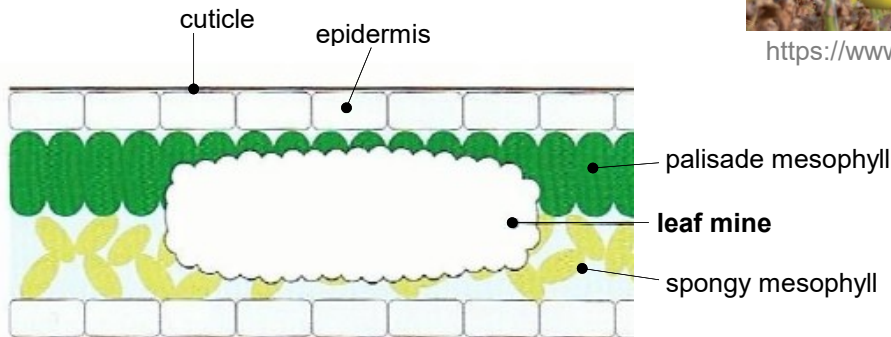
- Leaf mining caterpillars: feeding inside the leaf (e.g. *Tuta absoluta*)



<http://www.nbair.res.in/insectpests/Tuta-absoluta.php>

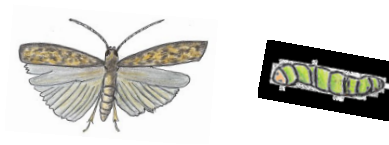


<https://www.bursadelegume.ro/influenta-atacului-tutei-absolute-asupra-pretului-tomatelor/>

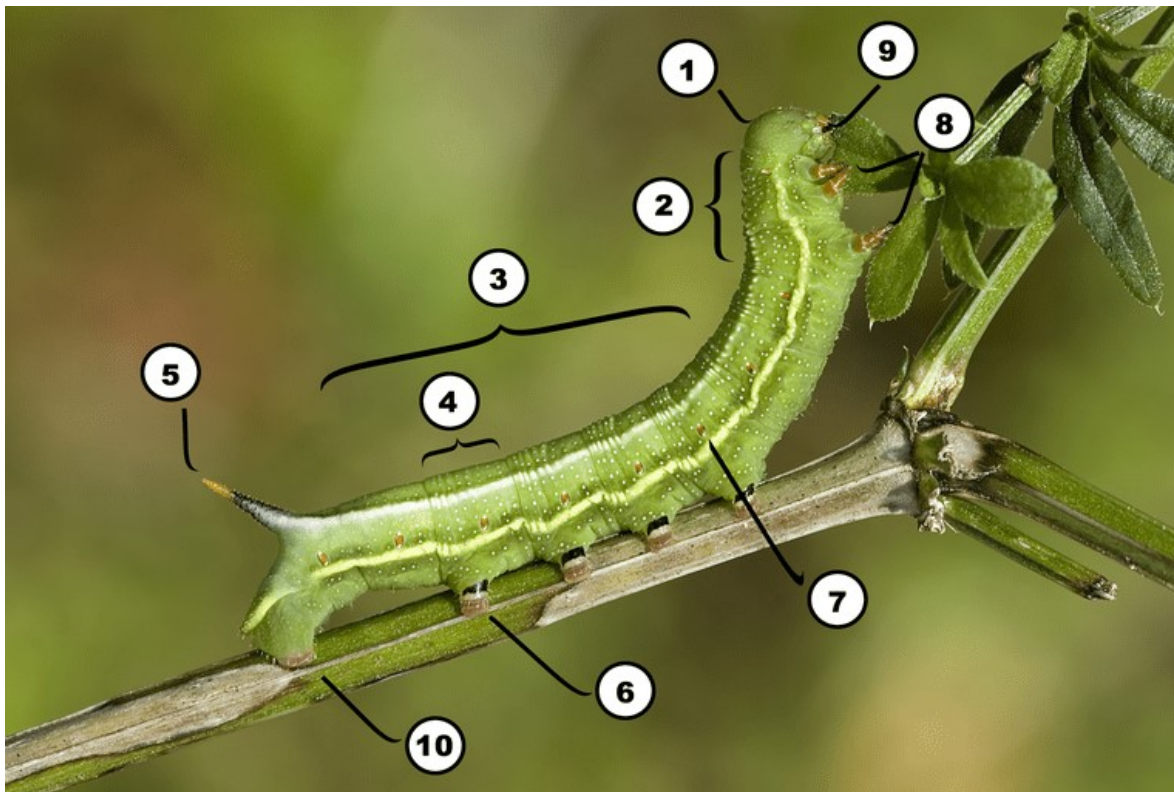


Double sided leaf mine (Jahrbücher des Nassauischen Vereins für Naturkunde, Band 130, Wiesbaden 2009)

Leaf miners feed inside the leaf, only minor damage to the epidermis. They can be a devastating pest, sometimes causing complete yield losses.



- Caterpillar anatomy

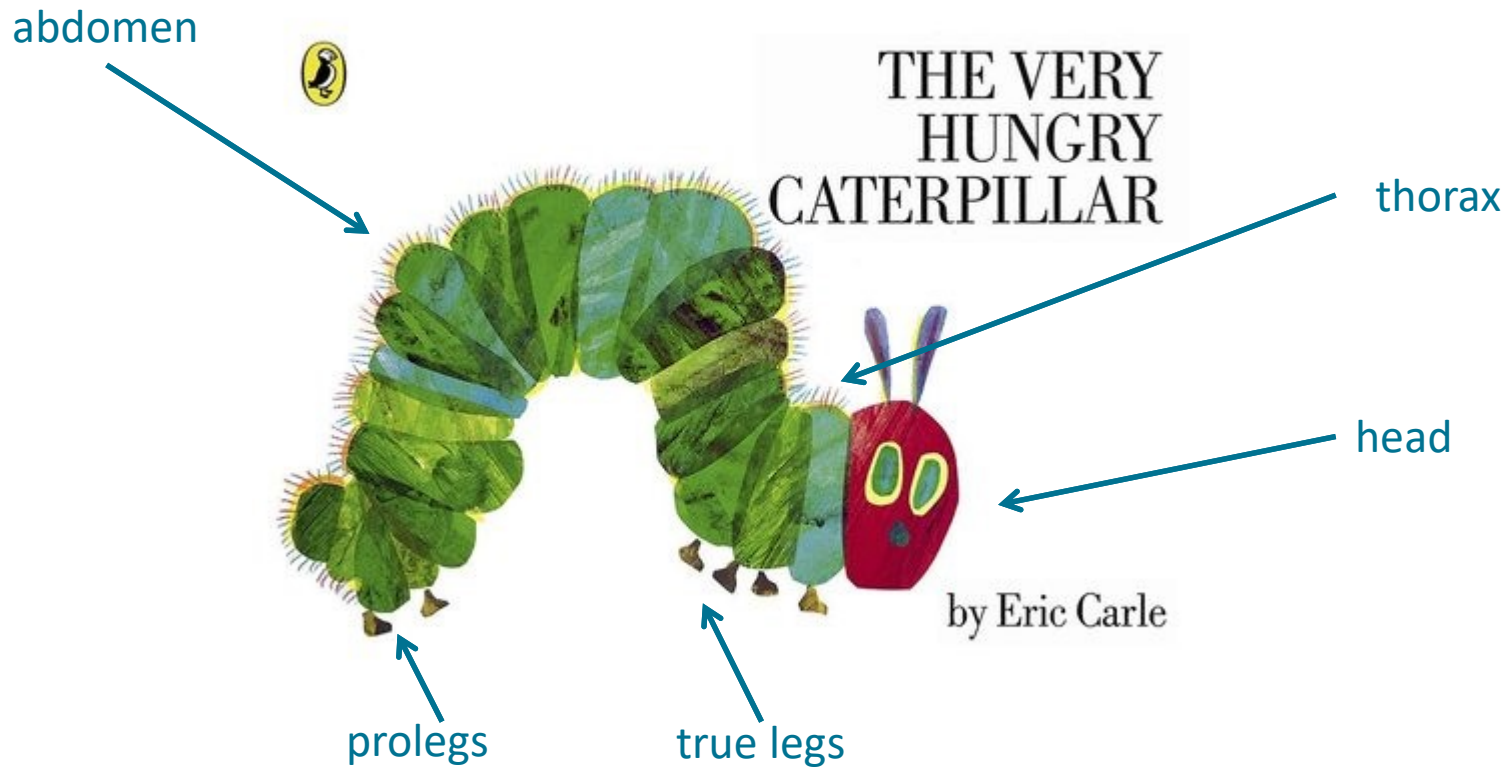


- 1: Head
- 2: Thorax with true legs (8)
- 3: Abdomen with prolegs (6; fleshy, false legs used to cling on, like 10: anal prolegs)
- 4: One segment
- 5: Horn (not with all species)
- 7: Spiracle: opening to respiratory system
- 9: Mandibles: jaws for chewing

A. M. Liosi/Wikimedia Commons (CC by SA license), modified by Debbie Hadley, WILD Jersey



- Caterpillar anatomy





- Caterpillar anatomy



AshLin

- **Mouth parts different from aphids**
 - different feeding mode, different kind of damage to the plant
- **Like aphids:** effectors in saliva; signal presence to plant (together with physical damage to leaves)



- Caterpillars: not all of them are ugly!



Ceruna vinula, the Puss Moth



- Caterpillars: not all of them are ugly!



Waugenberg

Calliteara pudibunda, Pale Tussock



- Caterpillars: not all of them are ugly!



Mjh233

Deilephila elpenor, Elephant Hawk Moth



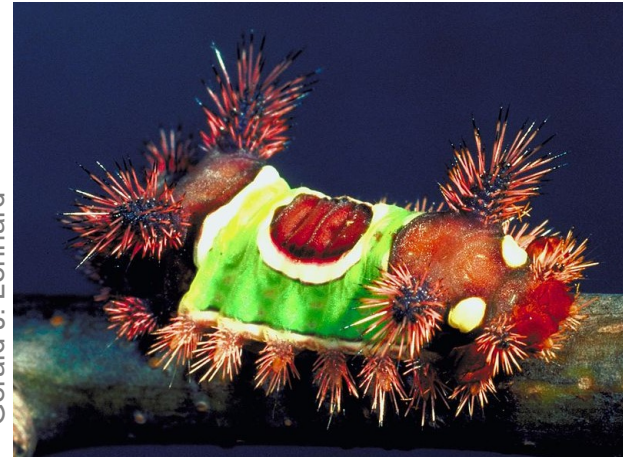
- Caterpillars: not all of them are ugly!

Georg Slickers



Orgyia antiqua, the Rusty Tussock

Gerald J. Lenhard



Acharia stimulea, the Saddleback caterpillar

Patrick Clenet



Aglais io, European Peacock

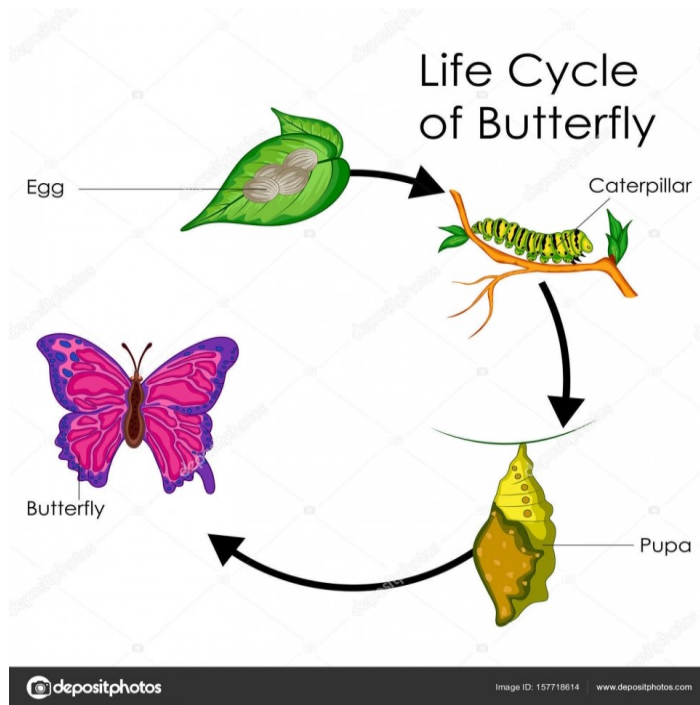
Gyorgy Csoka



Hypomecis roboraria, Great Oak Beauty



- Life cycle of caterpillars



All developmental stages of the Monarch Butterfly (*Danaus plexippus*)



- Life cycle of caterpillars: not all stages inflict damage on plants



Ferran Pestana

Adult Lepidoptera usually don't feed or only nectar, but lay eggs...



Andy Potter



Tony Wills

- **How can we control pests?**
 - Toxins: chemical insecticides
 - Biological control: natural enemies



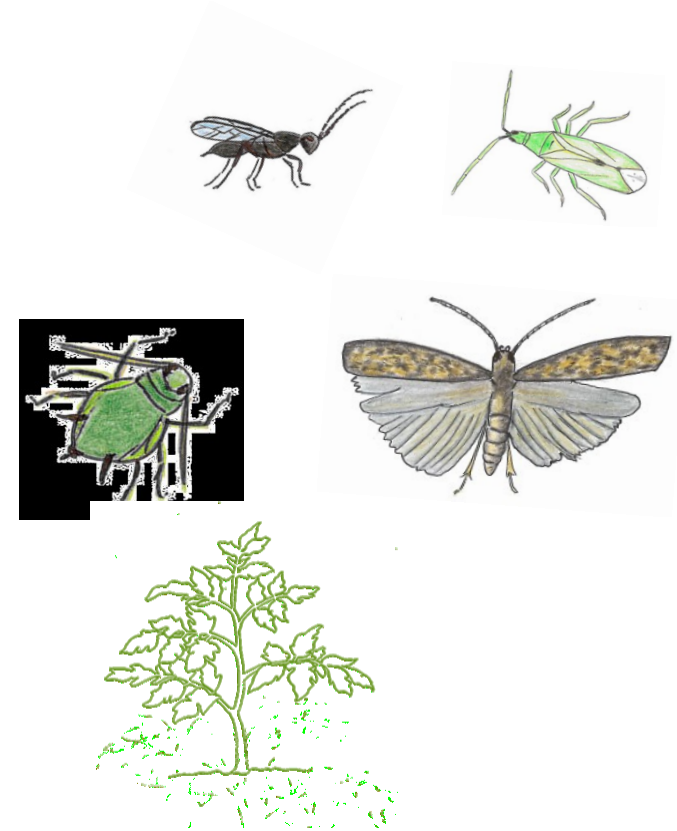
- Natural enemies: **Parasitoids**

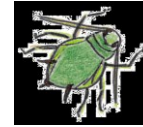
→ kill their hosts over time during own development



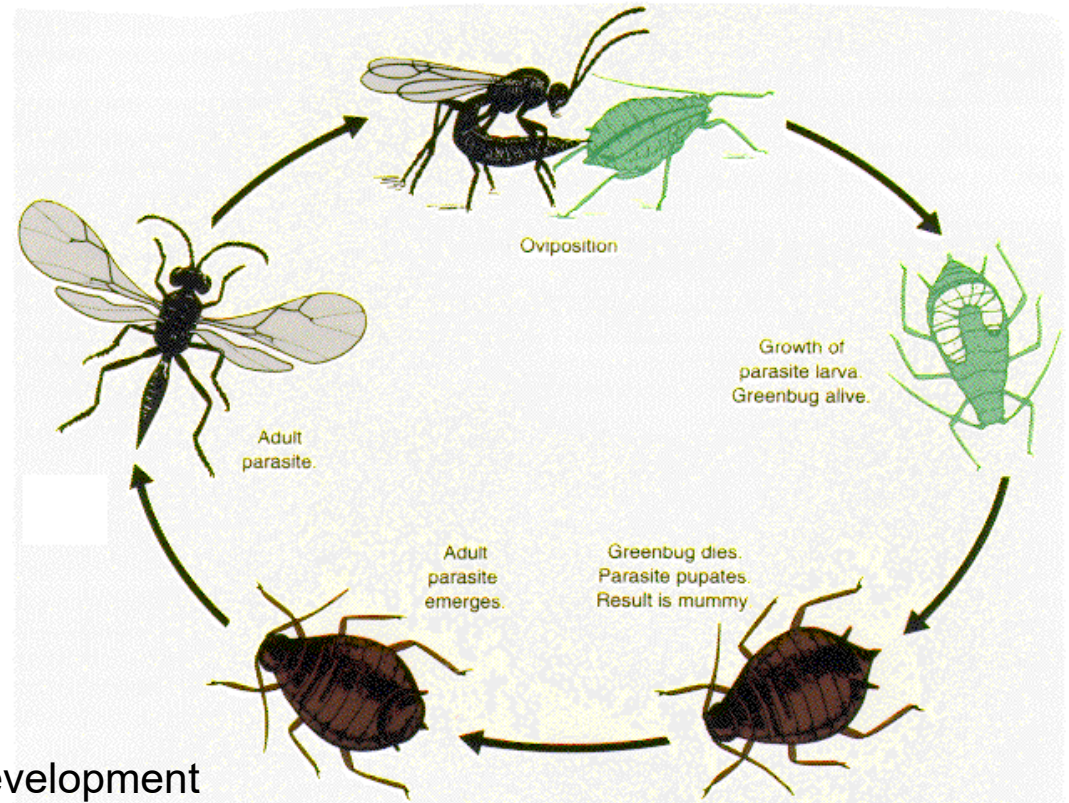
Alvesgaspar

A small braconid wasp laying an egg inside a Black Bean Aphid (*Aphis fabae*). The aphid is about 1 mm long.



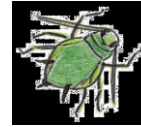


- Natural enemies: **aphid parasitoids**



Life cycle of an endoparasitoid:

- stings host, oviposits egg inside
- host still alive during offspring development
- offspring pupates, host dies
- adult parasitoid emerges



- Natural enemies: **aphid parasitoids**



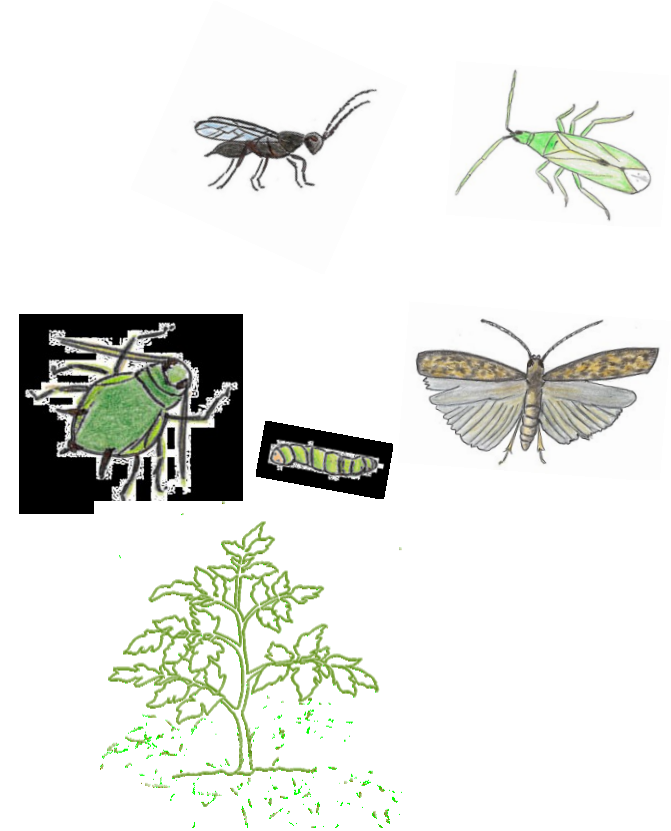
- for instance *Aphidius colemani*, *Aphidius ervi*
- one female lays 300-500 eggs in her lifetime
- wasp offspring develops from the egg to the adult inside the aphid
- parasitized aphids change color, become “mummies“, adult wasps emerge through round holes



- Natural enemies: **caterpillar parasitoids**



Aleiodes indiscretus wasp parasitizing a gypsy moth caterpillar (*Lymantria dispar dispar*)





- Natural enemies: **caterpillar parasitoids**

Ektoparasitoids: oviposit eggs on outside of host



Larvae of parasitic wasp *Bracon* sp.



Larvae of parasitic wasp
Cotesia congregata



- Natural enemies: **predatory bugs**

<https://www.entocare.nl/bestrijders/bestrijders-van-kevers/macrolophus-pygmaeus-feeding-on-pieris-brassicae-egg-1/>



The predatory bug *Macrolophus pygmaeus* feeding on eggs of *Pieris brassicae*

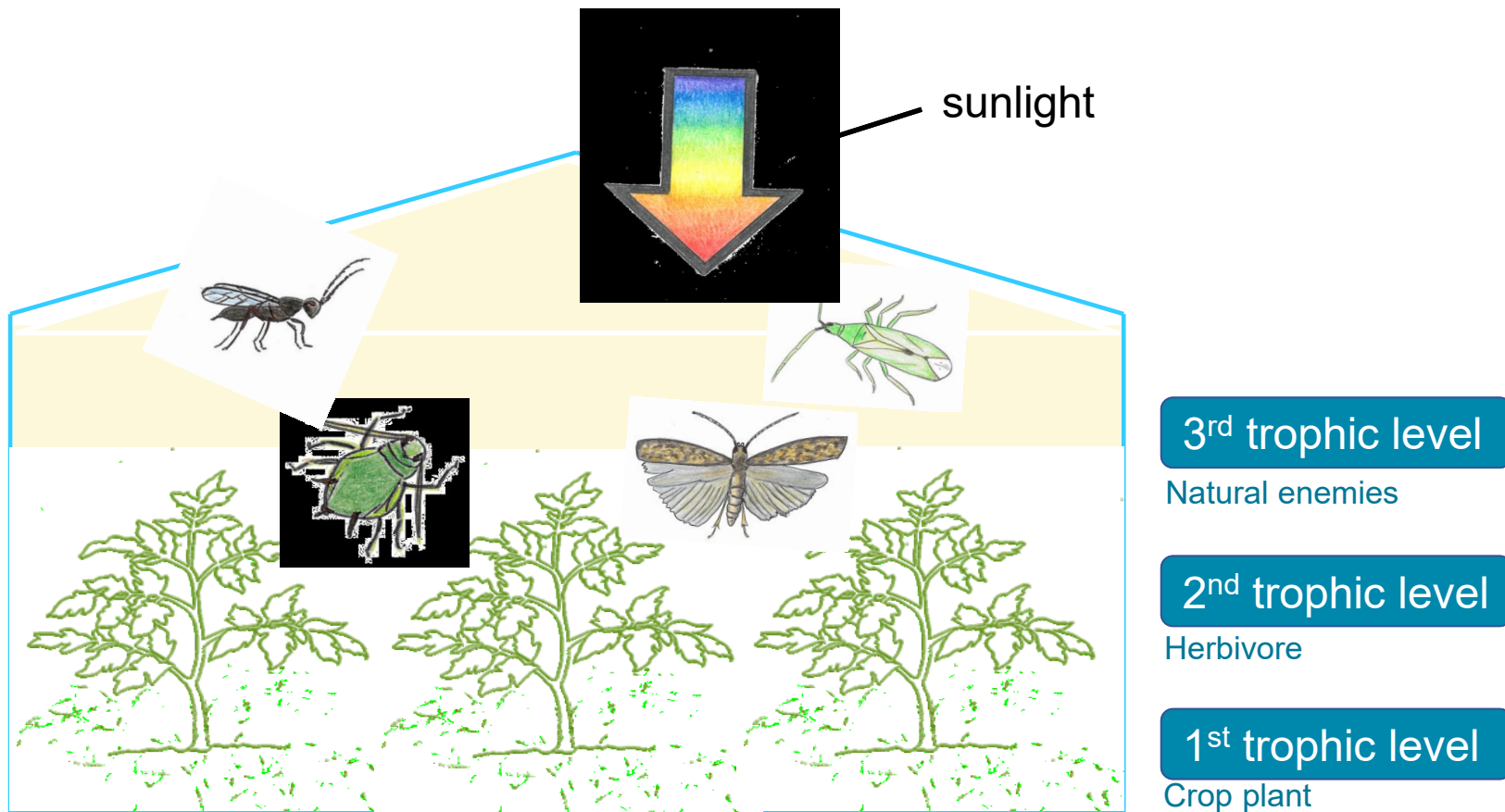
- Natural enemies: **humans?**



Pixabay

“Sago worm” satay

- Artificial ecosystem greenhouse: limited number of species
- but: trophic networks (“food chain“) exist, used for biological control



TROPHIC NETWORKS

Carnivores

3rd trophic level



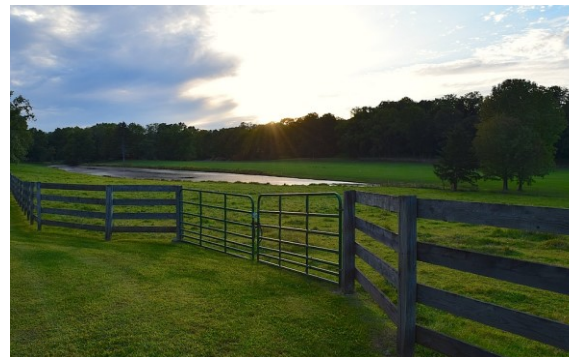
Herbivores

2nd trophic level



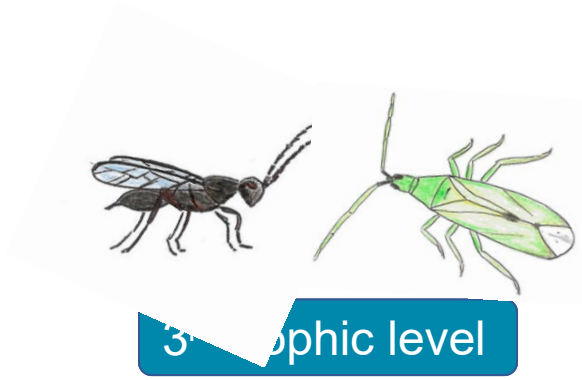
Plants

1st trophic level



TROPHIC NETWORK IN THE GREENHOUSE

Carnivores



- Natural enemies of pests: insect parasitoids and predators

Herbivores



- Herbivorous insects: leaf chewers like larvae of butterflies, and sap feeders like aphids and whiteflies

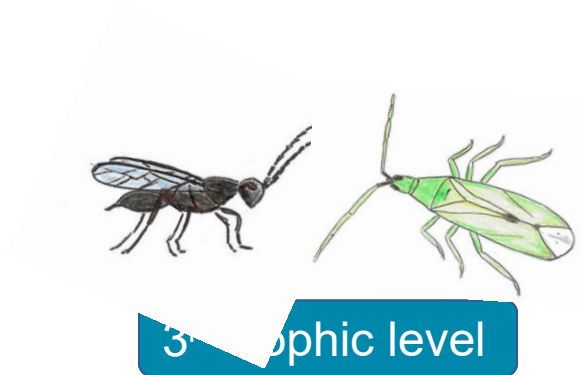
Plants



- Crop plants

DIRECT EFFECTS OF LIGHT

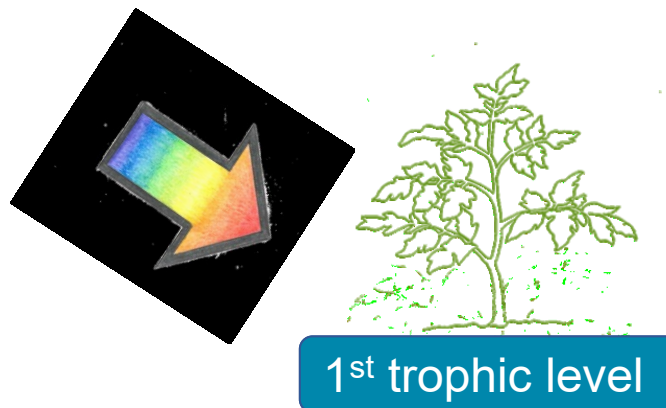
Natural enemies



Herbivores



Plants

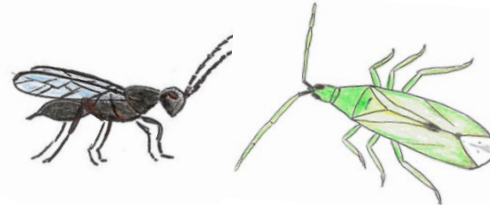


Direct effect:
on plant quality*

* nutritional value,
defense compounds

EXCURSION: PLANT DEFENSE

Natural enemies



3rd trophic level

Herbivores

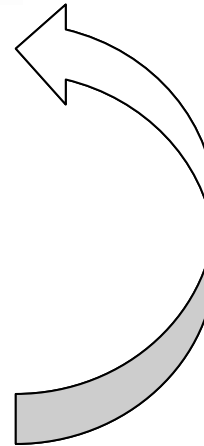


2nd trophic level

Plants



1st trophic level



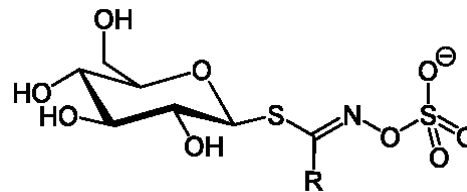
Direct defense:
morphology and
chemistry deter
herbivores

Direct defense:

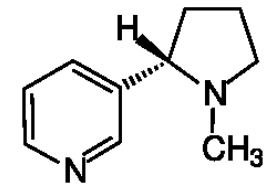
- Morphology: trichomes, thorns and spines, cuticula and epicuticular waxes, tough epidermis...
- Chemical defense: glucosinolates, alkaloids, phenolics...



<http://www.uva.nl/profiel/b/l/p.m.bleeker/p.m.bleeker.html>



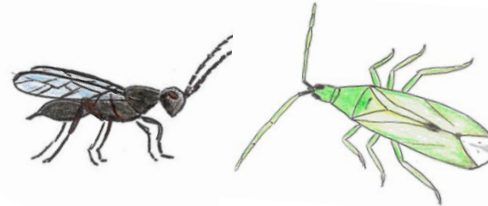
glucosinolate



nicotine
(alkaloid)

EXCURSION: PLANT DEFENSE

Natural enemies



3rd trophic level

Herbivores



2nd trophic level

Plants



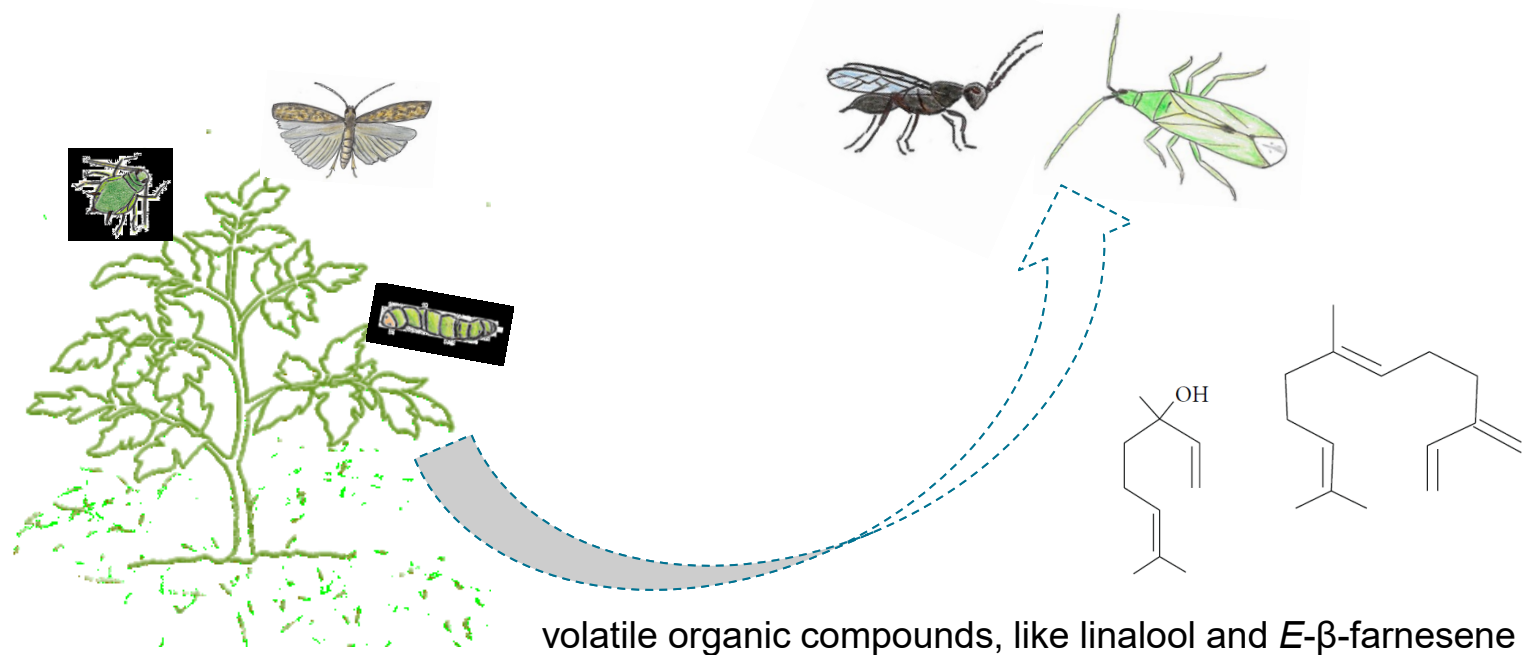
1st trophic level

Indirect defense:
e.g. VOC* attract
natural enemies of
herbivores

* volatile organic compounds

Direct defense:
morphology and
chemistry deter
herbivores

- **Indirect defense:**
- cellular food bodies and extrafloral nectaries as nourishment, domatia as shelter
- emission of VOC attracts natural enemies



What is the connection between plant defense and light conditions?

Plants are phototroph organisms.

→ light is the prerequisite for energy and carbon resources,
i.e. for every process from growth to defense!

...and if resources are limited?

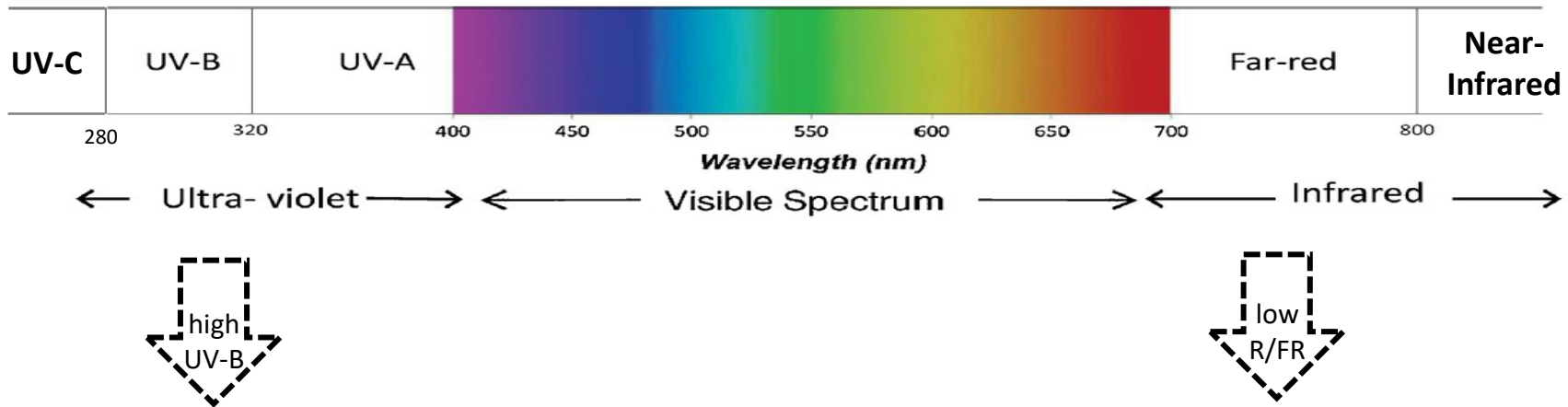
→ How to prioritize?

→ What is more important: growth or defense?

Well, that depends...



EFFECT OF LIGHT ON PLANT DEFENSE



growth < defense

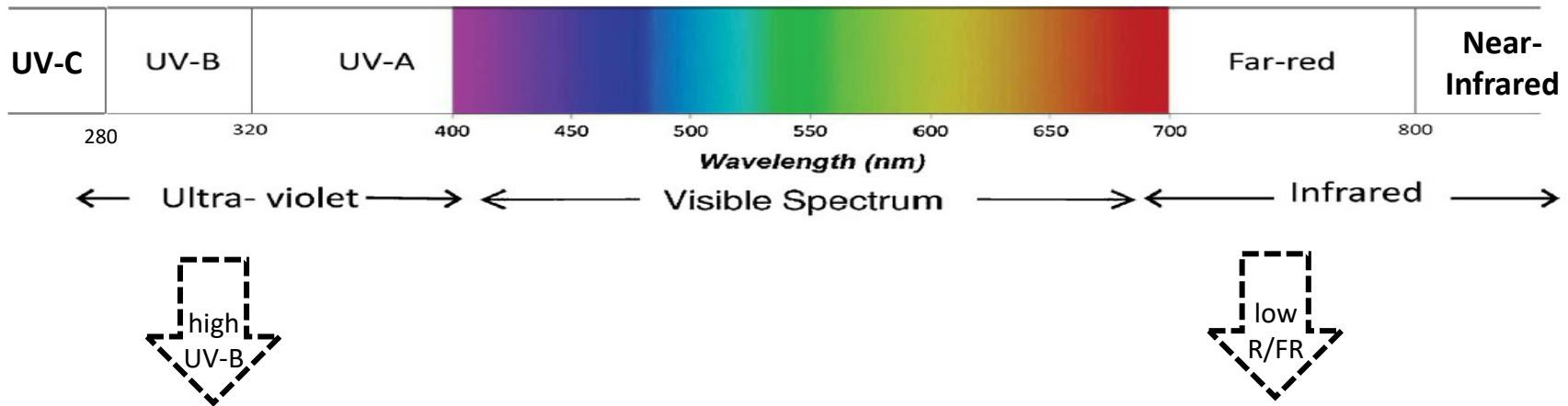
- High UV: plenty of light, no growth necessary, photosynthesis produces plenty of resources that can be invested in defense
- Can be a stress factor it self that warrants defense, dose dependency (interaction between biotic and abiotic stress when it comes to herbivory)



growth > defense

- shade avoidance-syndrome:
high FR: competition or shading, plants invest majority of resources into growth to reach canopy top
= foraging for light,
i.e. resources

EFFECT OF LIGHT ON PLANT DEFENSE



Probably well defended plants.

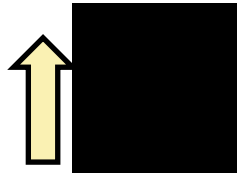


Probably weakly defended plants.

Huché-Thélier et al 2016, *Environ Exper Bot*, modified;
Ballaré et al 2009, *Plant, Cell Environ*; Ballaré & Perik 2017, *Plant Cell Environ*

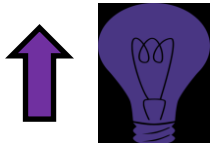
EFFECTS OF LIGHT ON PLANT DEFENSE

- light quantity:

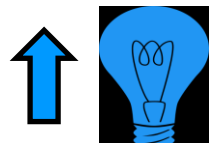


↑ trichome density and mechanical toughness of foliage,
chemical defense compounds (alkaloids, phenolics, terpenoids but not necessarily glucosinolates)
↑ nectar in extrafloral nectaries

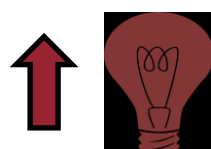
- light quality:



↑ UV-screening phenolic compounds, depending on intensity and frequency of exposure



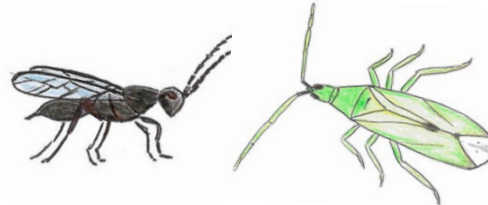
↑ blue light profoundly affects plant metabolism, including defence compounds, induces pathogenesis related-genes



↓ far-red reduced plant direct defense by desensitizing against herbivory-signaling,
decreased indirect defense (number of extrafloral nectaries)
↑ increased indirect defense (manipulated VOC → more attractive to predators; *few studies so far!*)

DIRECT EFFECTS OF LIGHT

Natural enemies



3rd trophic level

Herbivores



2nd trophic level

Plants



1st trophic level

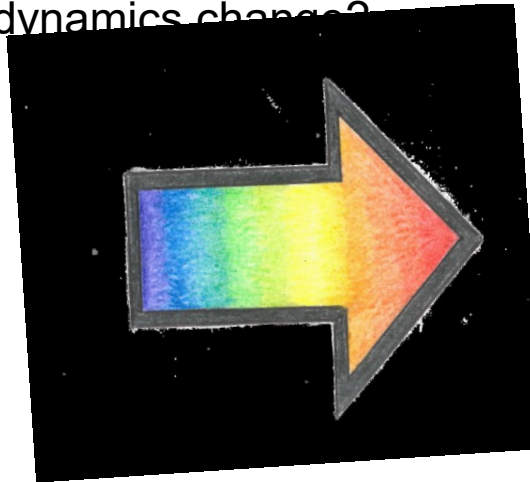
Direct effect:

on preference and
performance*

* orientation, host choice,
development

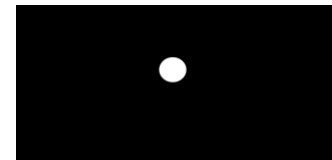
Light quantity and quality affects vision and orientation.

- Pest preference:
 - Can they find and recognize good host plants?
- Pest performance:
 - Can they find and recognize mates?
 - Do population dynamics change?



- What is necessary for a direct effect of radiation quality and quantity on herbivorous insects and their natural enemies?

PERCEPTION OF VISUAL CUES !



Eyes! (or something similar...)



- **Insects: several different organs to perceive light**
 - Ocelli (“simple eyes“)
 - Compound eyes

Picture: Wikipedia



dorsal ocelli



compound eyes



- **Ocelli:** “pigment pits“, simple eyes, one lense

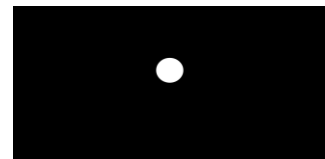


dorsal ocelli of a wasp



Picture: Wikipedia

dorsal ocelli of a hornet



human eye: “pigment pit“ with one lense



- **Compound eyes:**

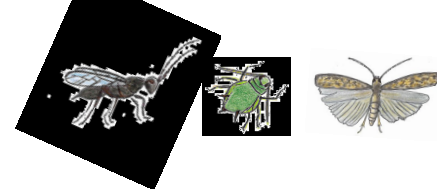


Compound eyes of a fly



Compound eyes of a robber fly

Picture: Wikipedia



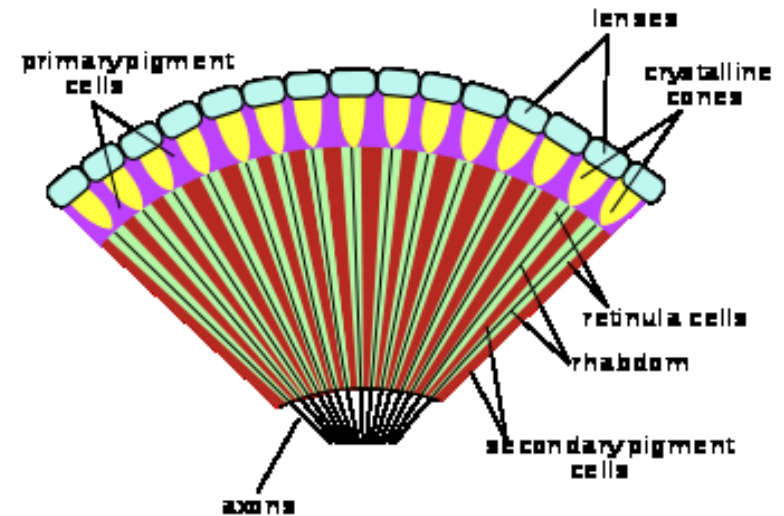
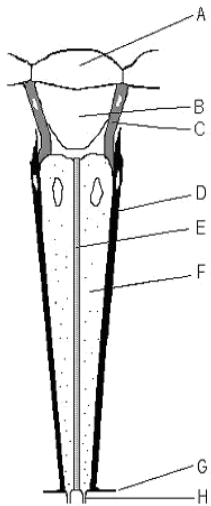
• Compound eyes:



Picture: Wikipedia

• Ommatidium: single eye

- cornea lense (A)
- crystal cone (B)
- pigment cells (C, D)
- rhabdom (E), guides light to the
- photoreceptor cells (F)
- membrane (G)
- optic nerve (H)

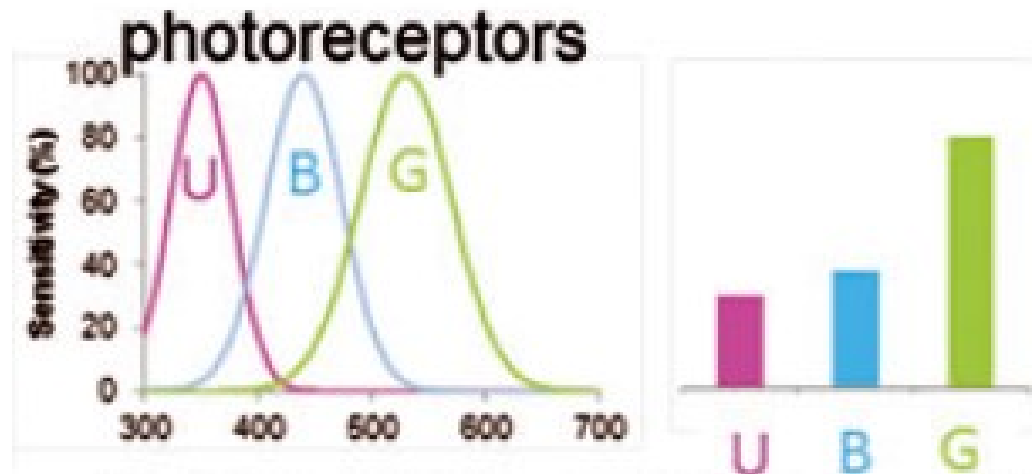


Wikipedia

- **Compound eye:** up to thousands of ommatidia



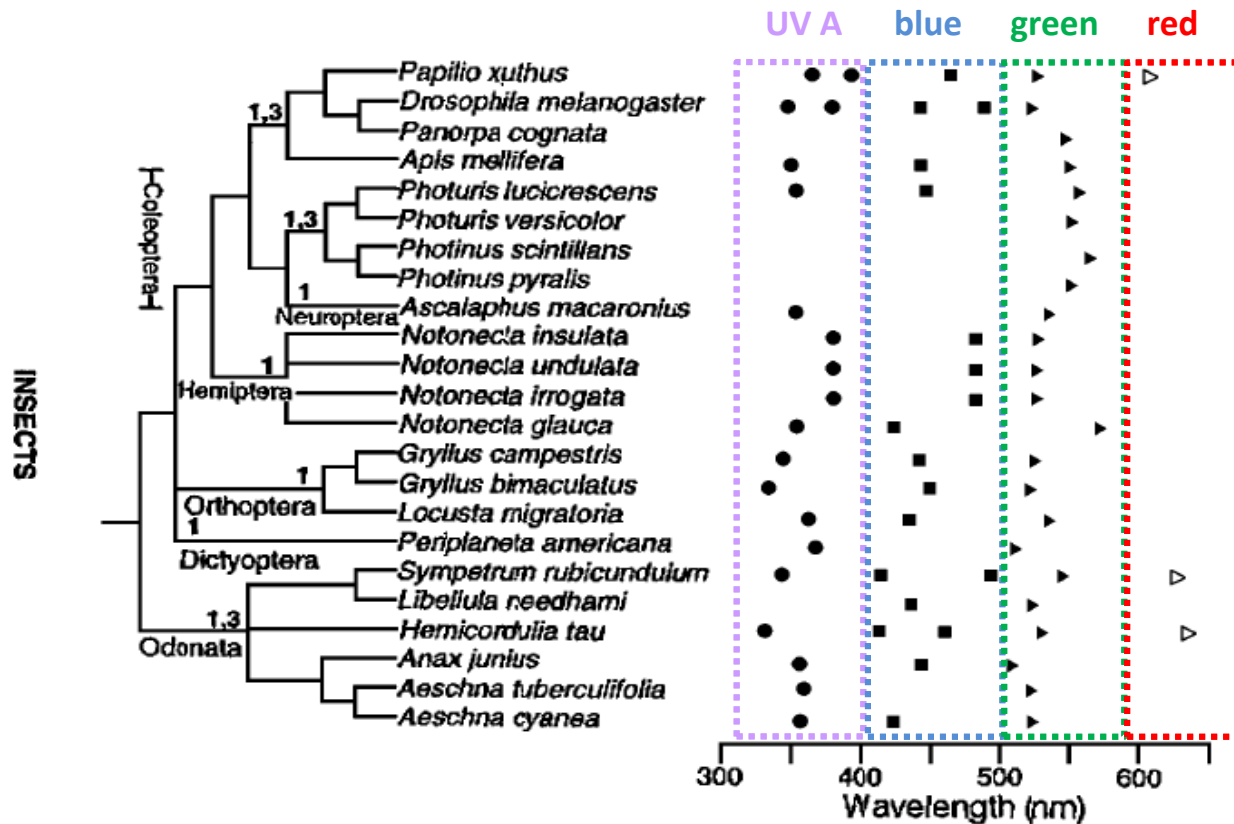
- The eyes contain photoreceptor cells located in the retina
- Photons are perceived if their wavelength is absorbed by the visual pigment:
opsin protein + chromophore; λ_{\max} = absorption maximum



Döring 2014, *Ann Appl Biol*

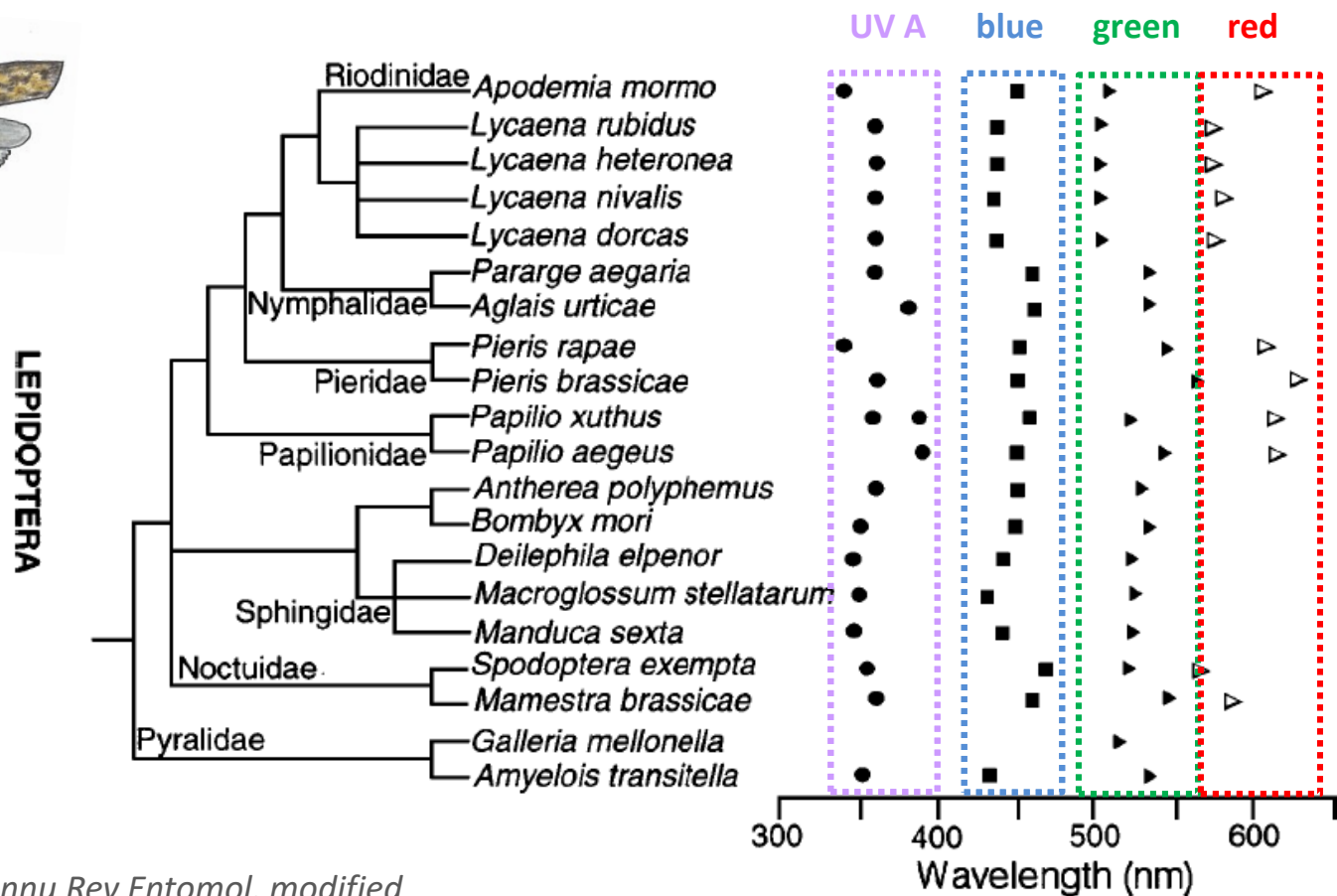


- Most insects perceive UV A (~350 nm) and green (~530 nm) wavelengths, many also blue (~440 nm), some also red (>565 nm)



PERCEPTION OF VISUAL CUES

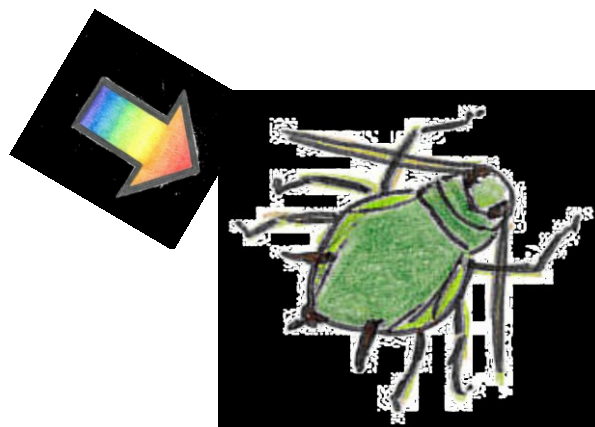
- Order of Lepidoptera: many well-studied and economically important herbivores



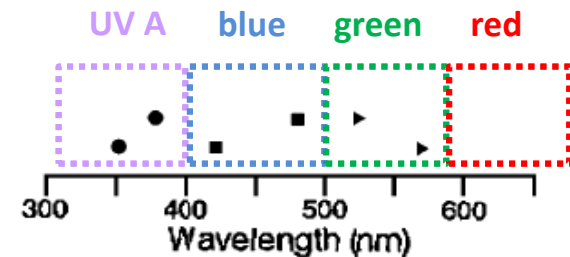
Briscoe & Chittka 2001, *Annu Rev Entomol*, modified

DIRECT EFFECT OF LIGHT ON PESTS

- Hemiptera*: mostly trichromatic as well (UV, blue, green)
 - for instance: whitefly, *Bemisia tabaci* and most aphids BUT some also perceive red
- Difficult to study due to small size of organs!



Hemiptera ——— *Notonecta irrogata*
 — *Notonecta glauca*



Briscoe & Chittka 2001, *Annu Rev Entomol*, modified

*Hemiptera: aphids, cicadas, planthoppers, leafhoppers, shield bugs

DIRECT EFFECT OF LIGHT ON PESTS

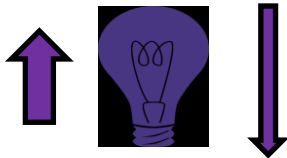
- **Light quality:**



orientation

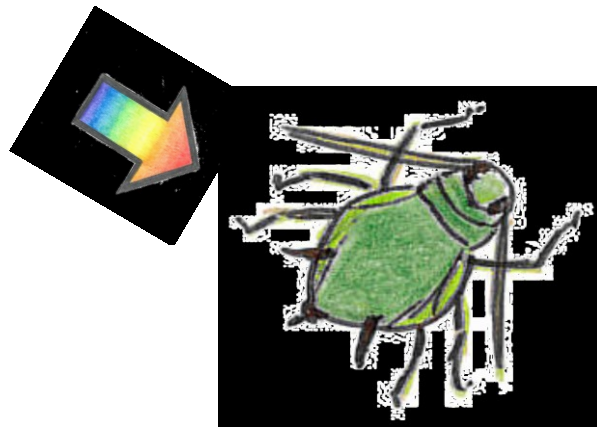


host plant location

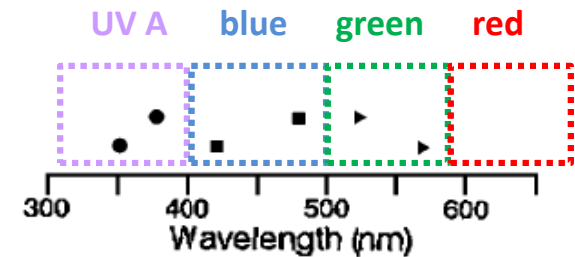


increased UV radiation reduced foraging and dispersal,
reduced survival of some pests

- **Response to wavelengths may differ with insect species & age!**



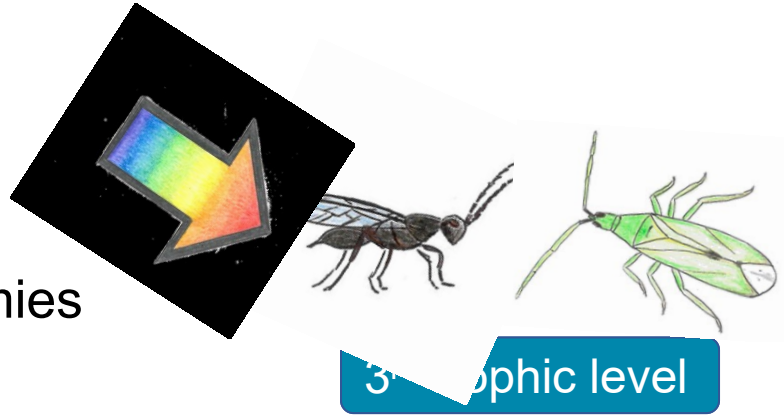
Hemiptera — *Notonecta irrogata*
 — *Notonecta glauca*



Briscoe & Chittka 2001, *Annu Rev Entomol*, modified

DIRECT EFFECTS OF LIGHT

Natural enemies



Direct effect:
on preference and
performance*

* orientation, host choice,
development

Herbivores

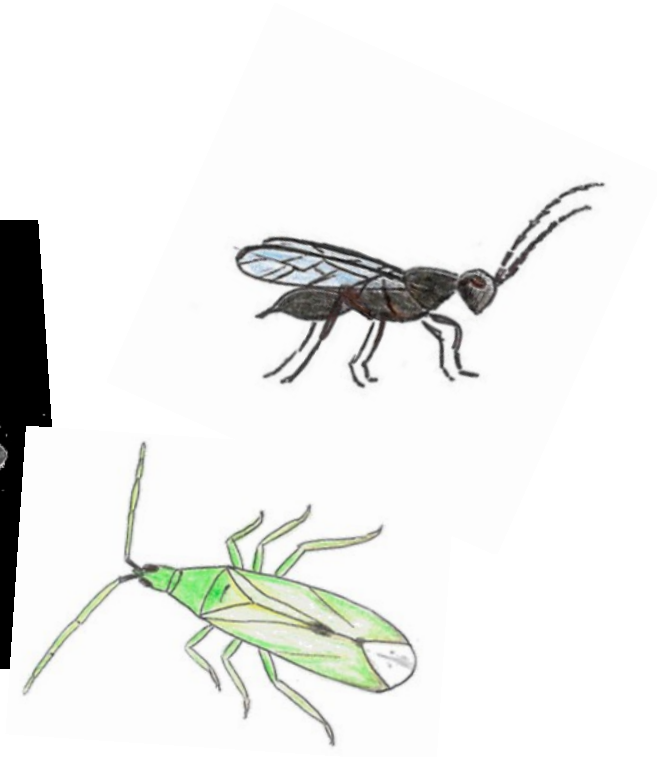
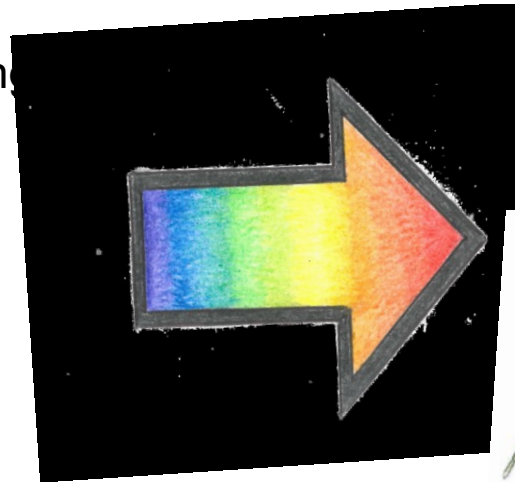


Plants

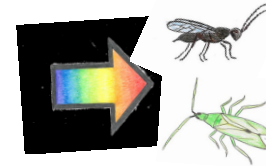


Light quantity and quality affects vision and orientation.

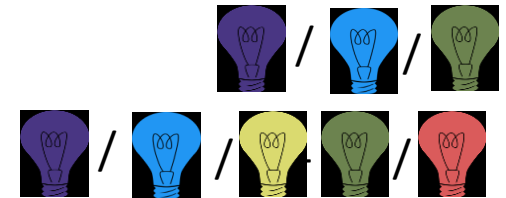
- Parasitoid preference:
 - Can they find and recognize good hosts?
- Parasitoid performance:
 - Can they find and recognize mates?
 - Do population dynamics change?



DIRECT EFFECTS OF LIGHT ON NATURAL ENEMIES

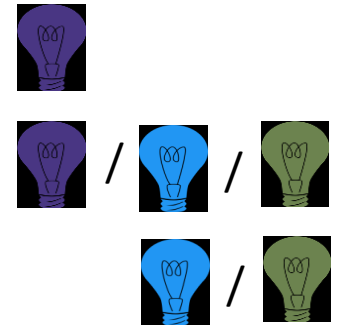


- **Vision of parasitoids:** similar to other insects; not as many studies...
- perceive mainly UV, blue, and green-yellow
- Parasitoid wasp, *Aphidius gifuensis*: UV, blue, green
- Parasitoid wasp, *Encarsia formosa*: UV, blue, green-yellow, red



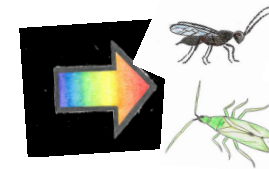
- **Phototaxis observed:**

- Tachinid parasitoid fly, *Exorista japonica*: towards UV-A
- *E. formosa*: towards UV-A, blue and green, but also active in UV-blocked environment
- Parasitoid wasp, *Scleroderma guani*: towards blue, green

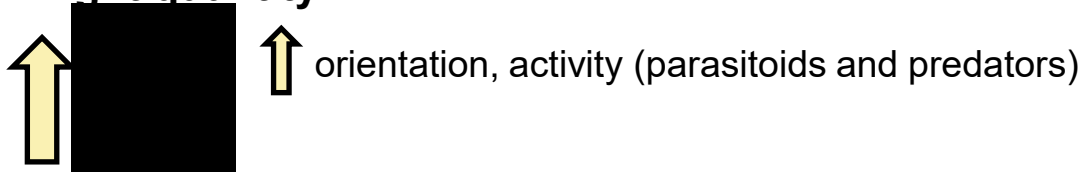


- **Response to wavelengths may differ with insect sex, age, and experience!**

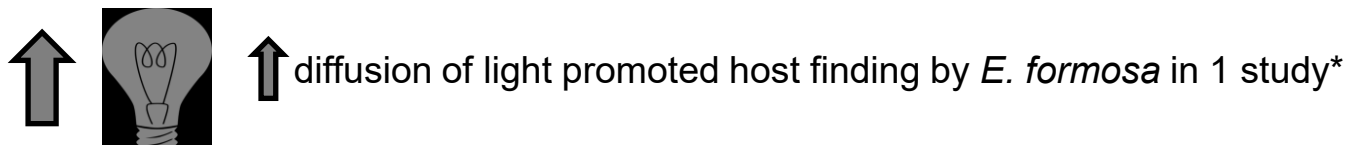
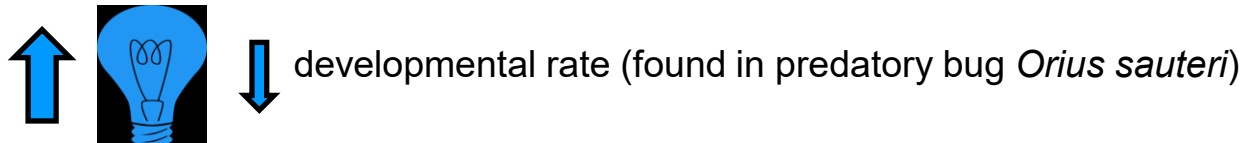
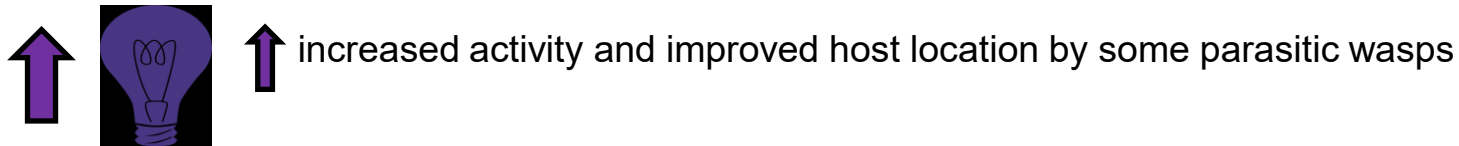
DIRECT EFFECTS OF LIGHT ON NATURAL ENEMIES



- **light quantity:**



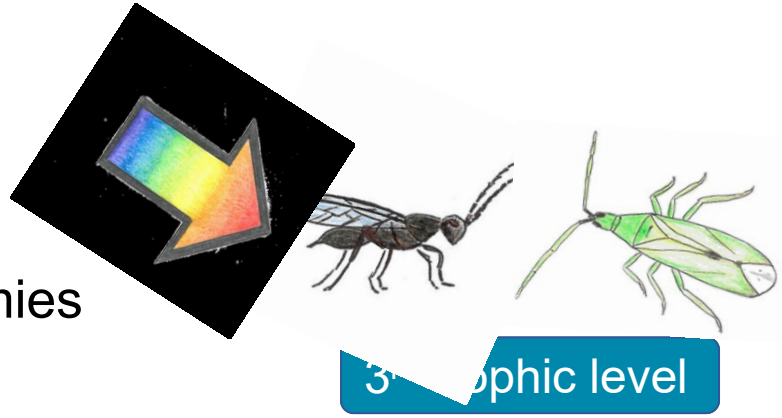
- **light quality:**



*Doukas & Payne 2007, *Horticultural Entomology*

TROPHIC NETWORK IN THE GREENHOUSE

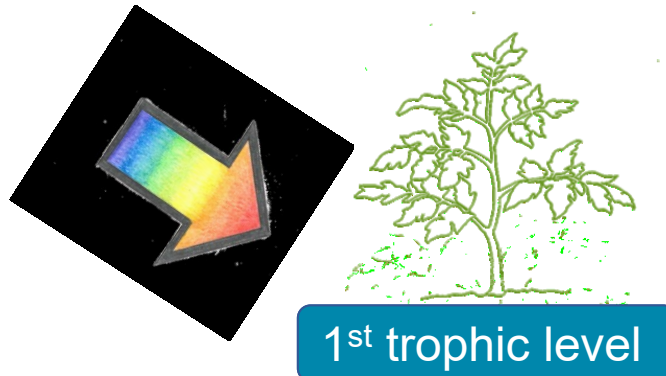
Natural enemies



Herbivores



Plants

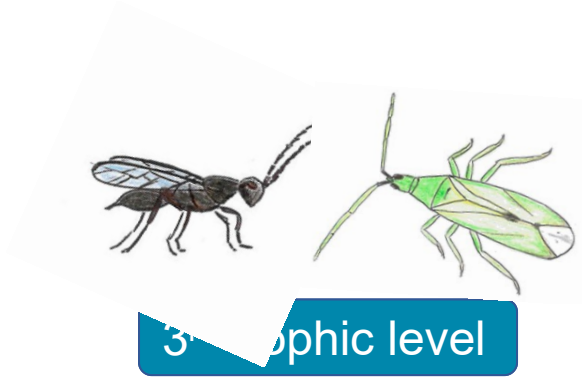


**Each trophic level
can be impacted
directly**

***But:* trophic
levels depend on
and interact with
each other!**

INDIRECT EFFECTS OF LIGHT: BOTTOM-UP

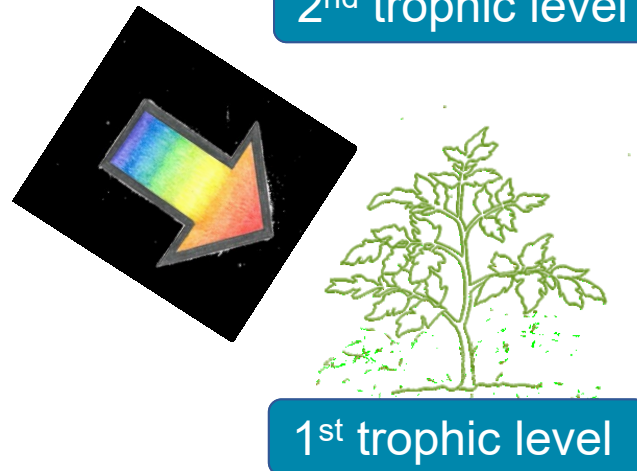
Natural enemies



Herbivores

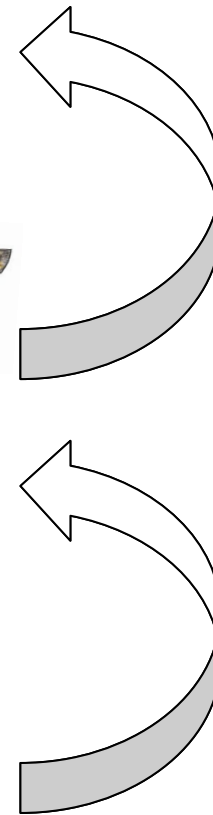


Plants



Bottom-up effect:
changes* induced
on 1st affect
higher levels

* nutritional value,
defense compounds



BOTTOM-UP EFFECTS

Carnivores

3rd trophic level



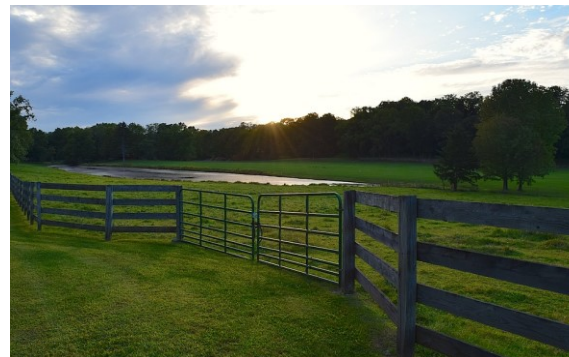
Herbivores

2nd trophic level



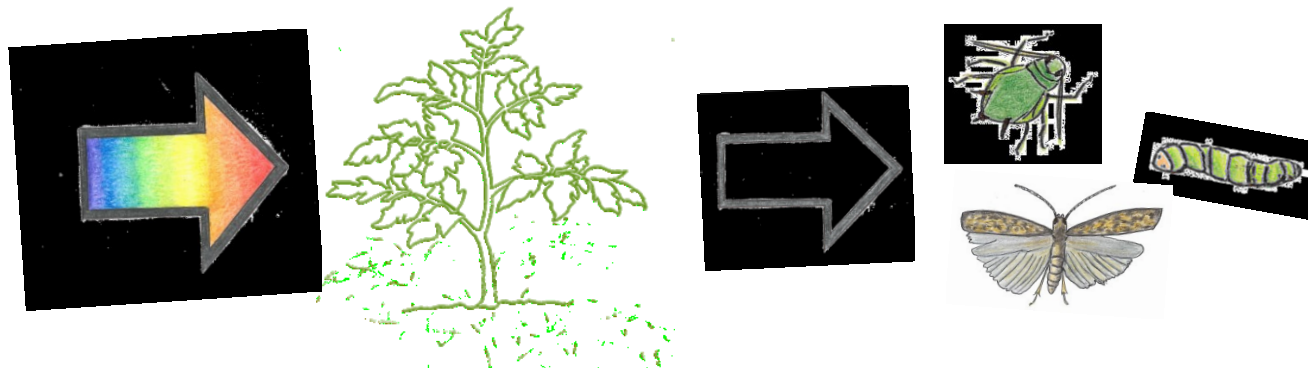
Plants

1st trophic level



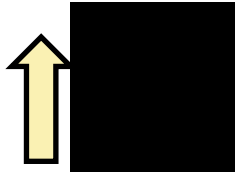
Light conditions affect plant quality. Pests consume/develop in/on plant tissue.

- Pest preference:
 - Can they find and recognize good host plants?
- Pest performance:
 - Does the plant offer enough nutrients for good development?
 - Has the plant accumulated toxic defense compounds?



INDIRECT EFFECTS OF LIGHT ON PESTS

- light quantity:

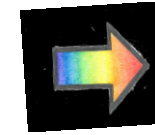


↓ decreased palatability of leaves due to enhanced toughness and defense chemicals (phenolics, terpenes, alkaloids)

↑ increased plant nutritional value, benefit for herbivores that can cope with also elevated defense compounds

↑ increased trichome density can trap more small arthropod pests (e.g. mites)

↑ increased VOC-emission; increased “chemical visibility“ to insects




→ In summary: probably lower infestation rates with higher light intensity


→ (But it's complicated: species, sex, development, experience...)

INDIRECT EFFECTS OF LIGHT ON PESTS

• light quality:



- ↑  ↓ reduces plant digestibility and attractivity for herbivores due to increased phenolics concentration → probably decreased herbivore performance
- ↑ flavonoid-rich larval diet enhances attractivity of adult females of *Polyommatus icarus* butterfly
- ↑ especially in the field, herbivores may benefit from UV-effect on entomopathogenic pathogens, and nematodes *

- ↑  ↑ increases preference of cabbage aphid (*Brevicoryne brassicae*) on broccoli **

→ In summary: probably higher infestation rates with lower UV radiation intensity.

→ But **it's complicated!** Herbivore and plant species, feeding mode and degree of dietary specialization are as important as experimental design...

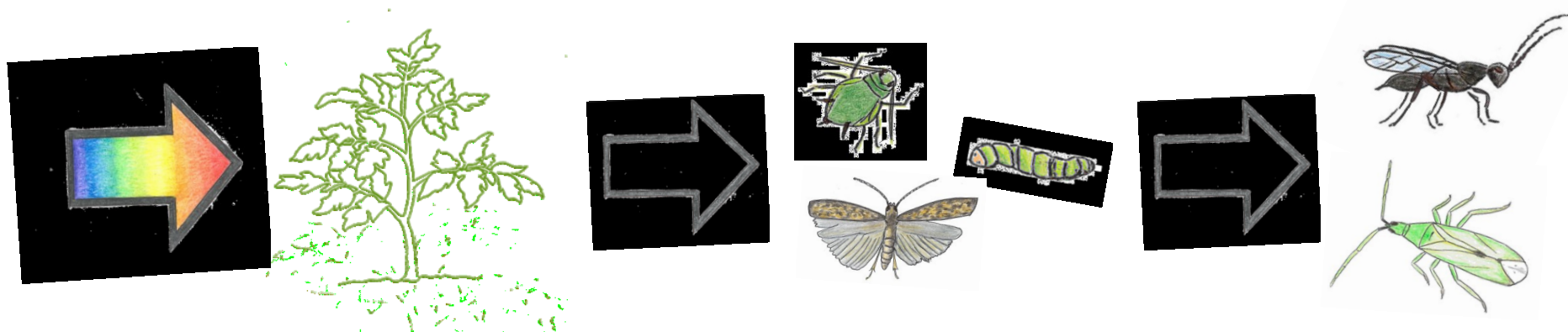
* Roberts & Paul 2006, *New Phytol*

**Rechner et al 2017, *Plos ONE*

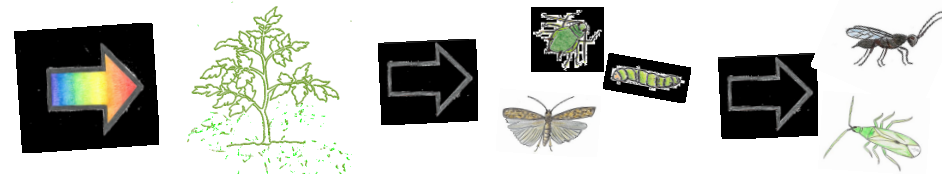
INDIRECT EFFECTS OF LIGHT ON NATURAL ENEMIES

**Light quantity and quality affect plant quality and therefore affect pests.
Natural enemies consume/develop in/on pest.**




- Natural enemy preference:
 - Can they find and recognize good insect hosts/ prey ?
- Natural enemy performance:
 - Does the insect host offer enough nutrients for good development and longevity?
 - Has the insect host taken up toxic plant defense compounds?






INDIRECT EFFECTS OF LIGHT ON NATURAL ENEMIES



• light quality:

   parasitic wasp *Cotesia plutellae* preferred diamondback moth larvae (*Plutellae xylostella*) that had fed on UV-B-exposed plants*

 No effect of elevated UV-B on *Cotesia marginiventris* and its host *Spodoptera frugiperda* **

   increased indirect defense (manipulated VOC → more attractive to predators; few studies so far!)

→ In summary: it depends on the involved species and experimental design.
(More studies needed.)



Invited Review

UV Radiation Effects on Pathogens and Insect Pests of Greenhouse-Grown Crops

Times Cited: 81
(from Web of Science Core Collection)

Michael Raviv¹ and Yehezkel Antignus²

¹Department of Environmental Horticulture, Neve Ya'ar Research Center, Agricultural Research Organization, Ramat Yishay, Israel

²Department of Virology, Volcani Center, Agricultural Research Organization, Bet Dagan, Israel

Received 6 June 2003; accepted 2 January 2004

ABSTRACT

Production of high-value crops is often performed under protected cultivation. In recent years various spectral modifications have been made in greenhouse covers. Two of the main reasons to modify the spectral characteristics of greenhouse covers have been to suppress the proliferation of several foliar diseases and to protect crops from insects. Insect-borne virus diseases of greenhouse-grown crops. These goals were achieved by complete or partial absorption of UV radiation, which interrupts the life cycle of several foliar pathogens and alters the visual behavior of many insects. Examples of these management strategies are described in this article.

Evidence from field studies. More than 80% of the studies that have evaluated the effects of ambient UVB on insect herbivory reported that plant damage or insect growth increased in response to attenuation of UVB radiation (10). Insect-feeding experiments with plants pretreated with either ambient or attenuated UVB radiation have clearly demonstrated that at least part of the effect of solar UVB reducing herbivory is indirect (i.e., mediated by changes in the quality of plant tissue). Ballaré 2014, *Annu Rev Plant Biol*

Contradiction?

et al. 2001; Kuhlmann and Müller 2009a). The species-specific and life-history trait-specific responses of herbivores to visual cues are probably driven by adaptations to different habitats. Under applied aspects, UV-absorbing filters can be used in integrated pest management programs at least against certain herbivorous insects (Antignus et al. 1996; Raviv and Antignus 2004).

Kuhlmann & Müller 2011, *Progress in Botany* 72

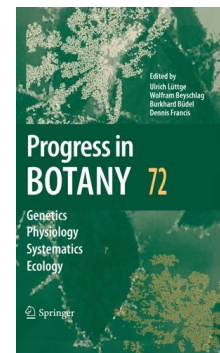
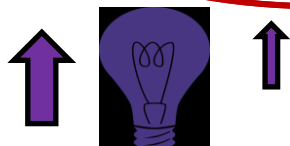
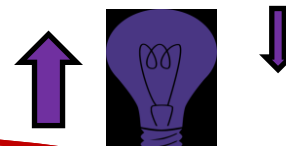
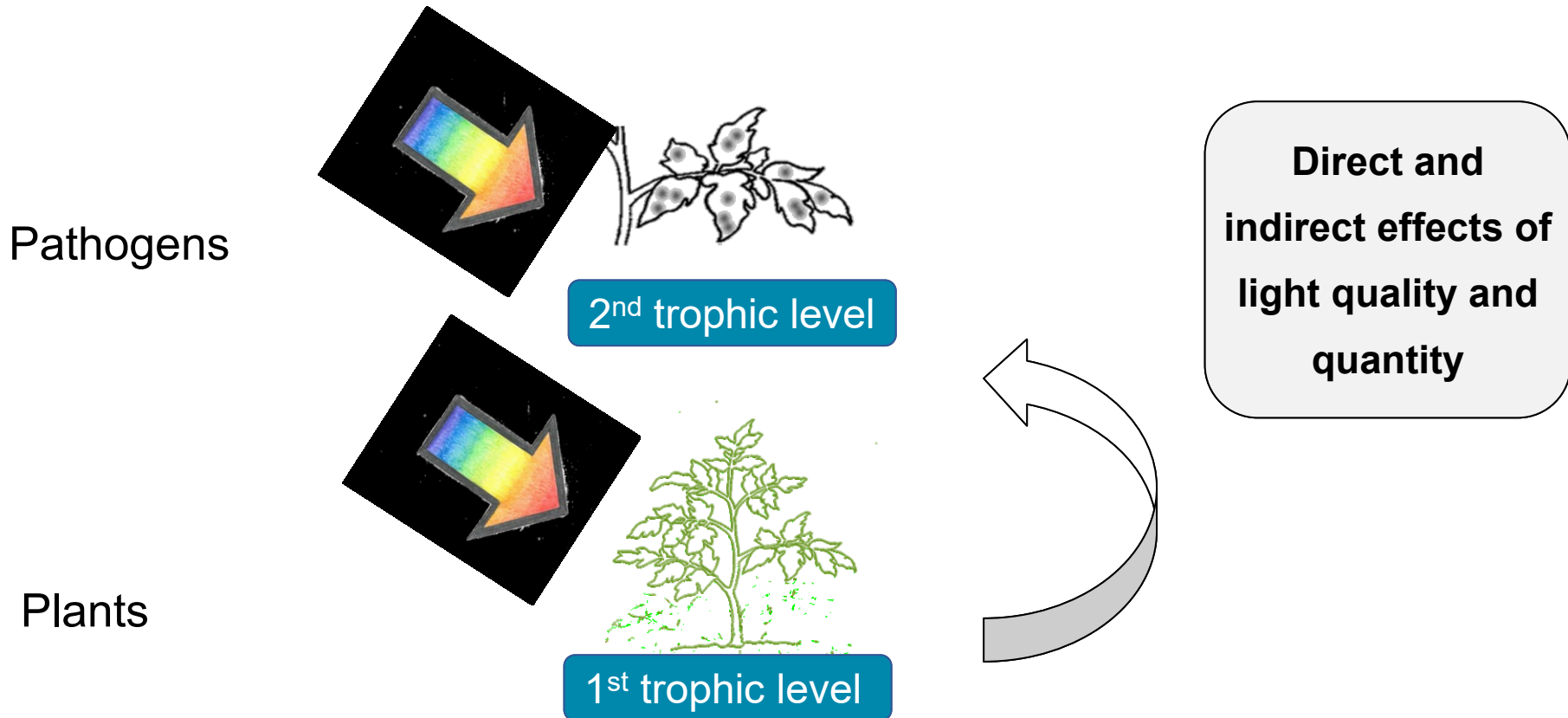


Table 1. Overview of ultraviolet (UV) radiation and its biological effects

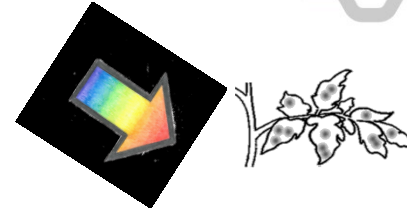
Waveband	Response to ozone depletion ^{a-c}	Physical and biological properties ^{a-c}	Role in animals ^{d-j}	Role in plants ^{d-j}
UV-C (200–280 nm)	None	Short wavelength, therefore highly energetic	None	None
UV-B (280–315 nm)	Absorbed in atmosphere; does not penetrate to biosphere Strongly affected by variation in the ozone column	Strongly absorbed by nucleic acids The most energetic UV radiation to reach biosphere	Artificial UV-C causes severe damage to skin and eyes Possible acute and chronic damage to skin and eyes	Artificial UV-C sources cause severe damage to exposed tissues Rarely damaging in the field but induces many morphological, physiological and biochemical changes in plants leading to many ecological effects
UV-A (315–400 nm)	Not affected	Absorbed by many biological molecules, including nucleic acids Less energetic than UV-B but present at much higher intensity in sunlight Absorbed by many proteins, including important photoreceptors	Compromises immune system Possible factor in decline of many amphibian populations Acute exposure causes sunburn in humans; chronic exposure is a cause of skin cancers Increasingly recognized as an important factor in development of skin cancers in humans Widely used in vision of many invertebrates and vertebrates	Influences plant morphology, plus some specific effects (e.g. stomatal opening and induction of 4pigment formation)

...AND WHAT ABOUT PATHOGENS?

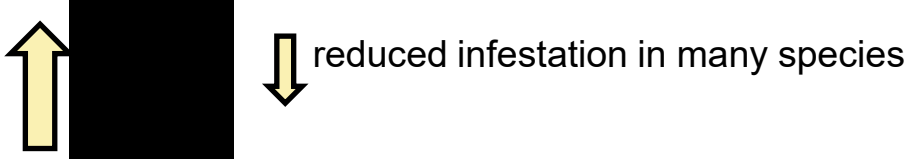
EFFECTS OF LIGHT ON PATHOGENS



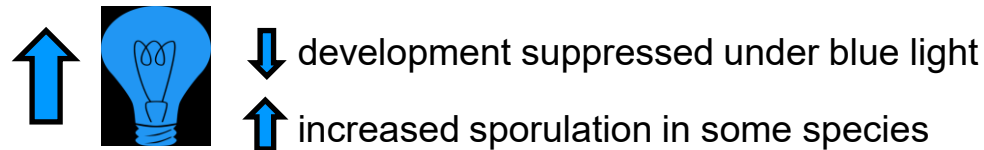
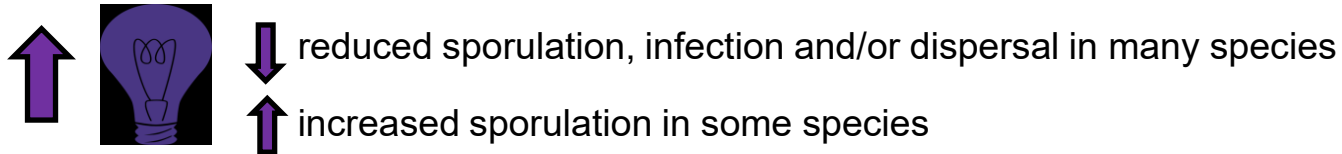
DIRECT EFFECTS OF LIGHT ON PATHOGENS



- **light quantity:**

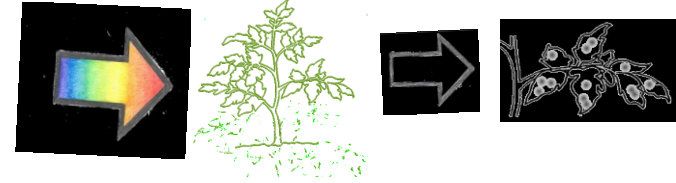


- **light quality:**

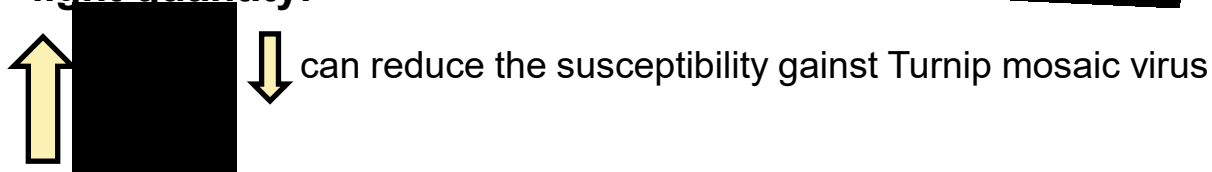


→ In summary: It depends on the involved species and their developmental stages

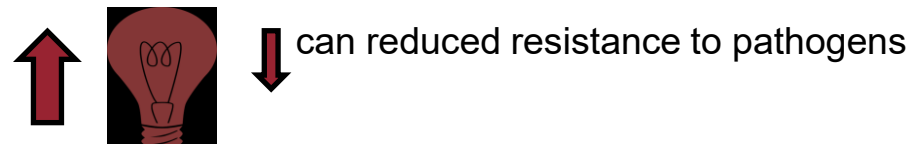
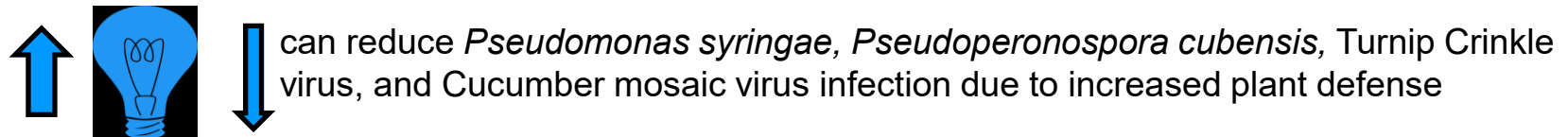
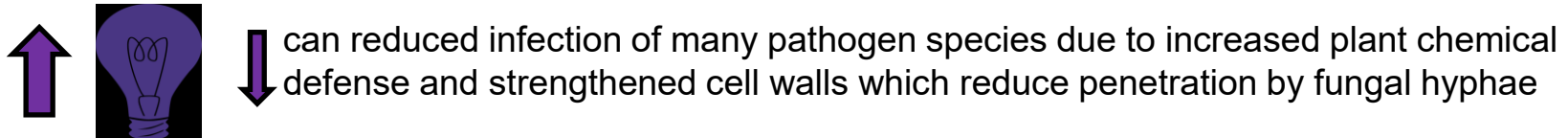
INDIRECT EFFECTS OF LIGHT ON PATHOGENS



- **light quantity:**

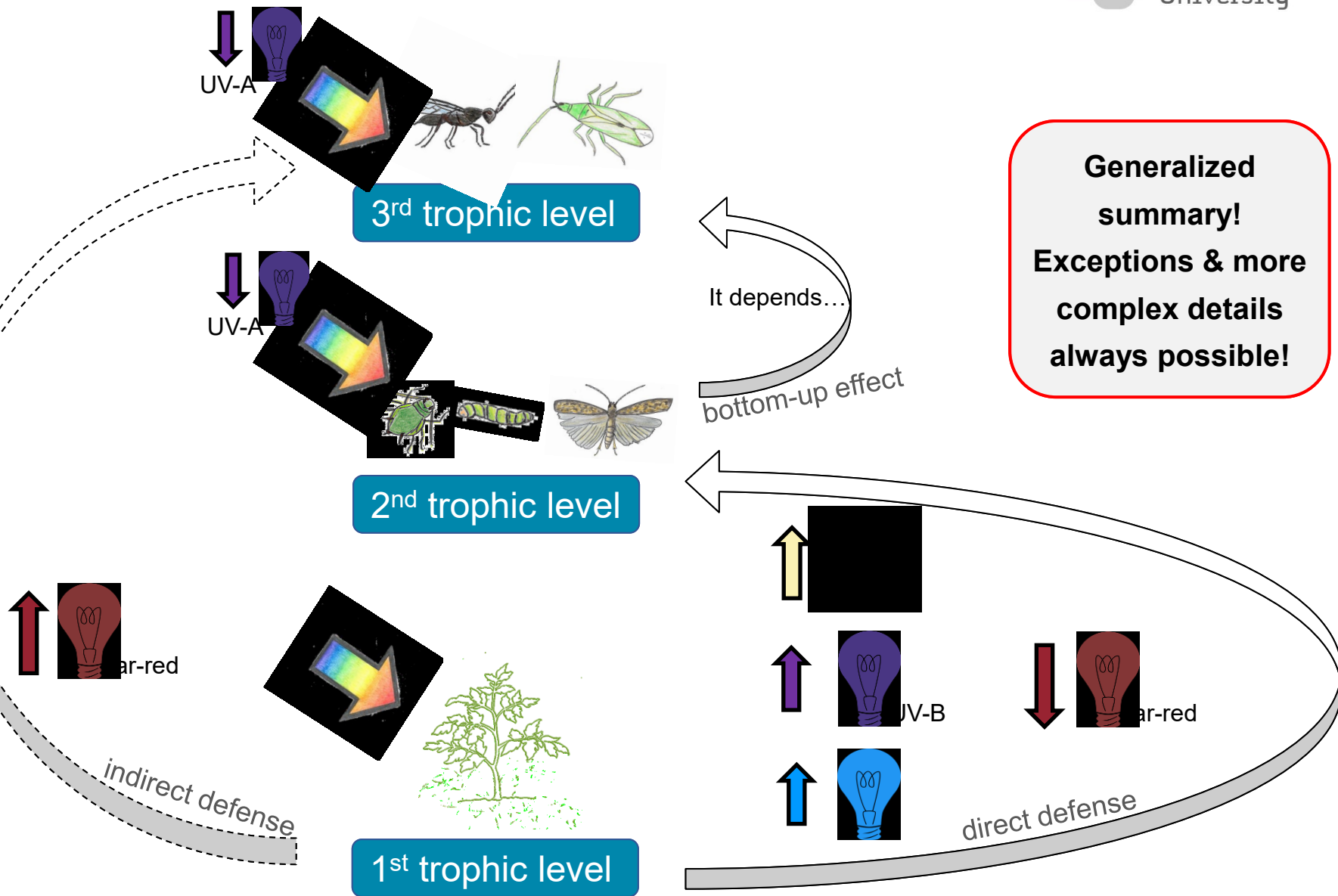


- **light quality:**



→ In summary: It depends on the involved species and their developmental stages

SUMMARY



THANK YOU FOR YOUR ATTENTION

Any questions?



Christine.Becker@hs-gm.de